X-ray and ¹H NMR Studies of the Conformational Equilibria of 2-Z-3-Phenyl-1,3,2-oxazaphosphorinanes. Steric and Stereoelectronic Influences on the Unexpected Axial Preferences of Me₂N and MeNH Substituents on Three-Coordinate Phosphorus

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A series of 2-Z-3-phenyl-1,3,2-oxazaphosphorinanes 7-14 (Z = MeO, (CF_3)₂CHO, Ph, MeNH, and Me₂N) containing three-coordinate phosphorus was prepared. The conformations of the six-membered rings were investigated by ¹H and ³¹P NMR spectroscopy and X-ray crystallography. The rings with substituents MeO, $(CF_3)_2$ CHO, Ph, and MeNH on phosphorus can be unambiguously assigned in solution to a single chair conformation with the substituent of phosphorus axial. An X-ray crystal structure of 5,5-dimethyl-2,3-diphenyl-1,3,2-oxazaphosphorinane, 11, reveals a chair form ring with the phenyl group attached axially to phosphorus. For 13 and 14 with a Me_2N substituent on phosphorus, a chair-chair equilibrium ($20 \Rightarrow 21$) is found in solution that features an 80-90% population $(\Delta G^{\circ} = 0.9-1.1 \text{ kcal/mol})$ of the Me₂N axial conformation (20). This finding contrasts sharply with the known 1 kcal/mol preference for the Me_2N to be equatorial in the corresponding 2-(dimethylamino)-1,3,2-dioxaphosphorinanes. The ability of the 1,3,2-oxazaphosphorinane ring to accommodate the Me2N substituent axially is also seen in the X-ray crystal of 5,5-dimethyl-3-phenyl-2-(dimethylamino)-1,3,2-oxazaphosphorinane, 13, which features a chair conformation ring that is considerably distorted compared to that of 11, quite evidently to allow the Me_2N to be in the observed axial orientation, conformation 20. It is argued that the axial orientation of the Me_2N in 13 and 14 is at least partly in response to steric repulsions in the alternative chair conformation 21 between the equatorial Me_2N and the phenyl substituent at N(3). This effect is *in direct contrast* to the repulsive interactions between the N(3)Ph and axial Me₂N on phosphorus previously demonstrated for *four-coordinate*, 2-oxo-1.3.2-oxazaphosphorinanes. The increased bond lengths within the 1.3.2-oxazaphosphorinane ring over its 1,3,2-dioxaphosphorinane counterpart (C-N vs C-O) and increased ring flexibility, along with potential $n \rightarrow \sigma^*$ stereoelectronic factors of the type operative in the anomeric effect, are also proposed as potential contributors to the preferred axial orientation of Me₂N in 13 and 14. The diastereomeric molecules cis- and trans-5-tert-butyl-3-phenyl-2-(dimethylamino)-1,3,2-oxezaphosphorinane, 17, also were prepared. At thermodynamic equilibrium at room temperature, cis-17 $(2-Me_2N \text{ and } 5-t-Bu \text{ groups cis})$ is favored (cis/trans = 80/20). cis-17 displays a conformational equilibrium (Scheme 1) involving a chair conformer ($\approx 60\%$) with the t-Bu equatorial and Me₂N axial, cis-17a, and a single twist or boat form with both substituents pseudoequatorial, cis-17d ($\approx 40\%$). trans-17 exists in solution in three conformations in approximately equal populations: a chair form with both t-Bu and Me₂N equatorial (trans-17a) and two boat/twist forms (trans-17b and trans-17c) with the t-Bu pseudoequatorial and the Me₂N pseudoaxial. The distributions of chair and boat/twist conformations can be reasonably understood in terms of the same 1,3-syn axial and vicinal PhN- $(3)/Me_2N(eq)$ steric repulsions invoked to explain the chair-chair equilibria noted for the unsubstituted and 5,5-dimethyl-2-(dimethylamino)-3-phenyl-1,3,2-oxazaphosphorinanes 13 and 14. The free energy difference between chair and boat/twist forms evidently is very small.

Introduction

The conformational properties of heterocyclic sixmembered rings containing O, N, S, and P atoms have been the subject of considerable study.¹ Most thoroughly investigated have been the conformations of both threeand four-coordinate 1,3,2-dioxaphosphorinanes. Substituents on three-coordinate phosphorus show a preference for the axial position in the order ArO > RO > Ph > Me > *i*-Pr. The dimethylamino group and the sterically large *t*-Bu prefer the equatorial orientation.¹ MeNH displays an intermediate, more nearly neutral, axial-equatorial preference.¹ The conformations of four-coordinate 2-oxo-1,3,2-oxazaphosphorinanes, which include the important antitumor drugs cyclophosphamide, 1, and its congeners trofosphamide, 2, and isofosphamide, 3, also have been examined.² The nature of the substituent on N(3) has been found to be of great importance in determining the orientations of more bulky groups such as Me₂N on phosphorus. A major effect of ring heteroatoms is the lowering of the free energy differences between chair and twist conformations as is observed for both 1,3,2-dioxaand 1,3,2-oxazaphosphorinanes.³ Thus, the conformational energies of six-membered rings containing heteroatoms are strongly influenced by steric interactions and by stereoelectronic effects.²

Surprisingly, the conformations of three-coordinate 1,3,2-oxazaphosphorinanes have received little attention. Although a series of such compounds was prepared by Nifant'ev et al.,⁴ a detailed conformational analysis was

[•] Abstract published in Advance ACS Abstracts, October 1, 1993. (1) Reviews may be found in: (a) Bentrude, W. G.; Setzer, W. N. In Phosphorue-31 Spectroscopy in Stereochemical Analysis, Verkade, G., Quin, L.D., Eds.; VCH Publishers: Deerfield Beach, FL, 1987; pp 365-389. (b) Maryanoff, B. E.; Hutchins, R. O.; Maryanoff, C. A. Top. Stereochem. 1979, 11, 187-317.

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not attempted. However, on the basis of ¹H and ¹³C NMR data, Nifant'ev et al. concluded that 2-(dimethylamino)-3-isopropyl-1,3,2-oxazaphosphorinane (4), 2-phenyl-3-isopropyl-1,3,2-oxazaphosphorinane (5), and 2-ethoxy-3isopropyl-1,3,2-oxazaphosphorinane (6) all exist in chair conformations with substituents on phosphorus axial (20).^{4a} In fact, as we will show, 1,3,2-oxazaphosphorinane 4 is 75–80% in this conformation. This 0.7 kcal/mol axial propensity of the 2-Me₂N group is surprising, since the Me₂N group displays a 1 kcal/mol preference for equatorial attachment to phosphorus in three-coordinate 2-(dimethylamino)-1,3,2-dioxaphosphorinane.⁵

$$4 Z = NMe_2$$

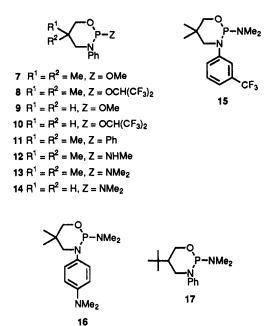
$$5 Z = Ph$$

$$6 Z = OE1$$

We reported preliminary results of the conformational study of a series of 3-phenyl-1,3,2-oxazaphosphorinanes.⁶ The more highly populated conformer for 13 in that series was the chair form with the 2-Me₂N axial (20, $H_Y = H_X = CH_3$). In the present paper, we examine the chair-chair equilibria, corresponding to $20 \Rightarrow 21$ of three-coordinate 3-phenyl-1,3,2-oxazaphosphorinanes 7-16 in detail. Like the corresponding three-coordinate 1,3,2-dioxaphosphorinanes, 7-11 are in a single chair conformation, 20 (or the 5,5-dimethyl analog).

Thus, MeO, $(CF_3)_2$ CHO, and Ph substituents on phosphorus display a strong axial preference. However, *unlike* the three-coordinate dioxaphosphorinanes, 12–16 also are very largely in chair conformations with the MeNH or Me₂N group axial in agreement with Nifant'ev's finding

(6) Huang, Y.; Mullah, N. N.; Sopchik, A. E.; Arif, A. M.; Bentrude, W. G. Tetrahedron Lett. 1991, 32, 899-902.



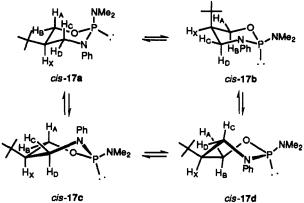
for 4. Furthermore, the chair-chair equilibrium for threecoordinate oxazaphosphorinanes 13-16 is seen to be not significantly influenced by solvent change or placement of either electron-withdrawing or -donating groups on the phenyl substituent at N(3). Lastly, we report a study of the chair-twist equilibria for previously unreported *cis*and *trans*-17 (Schemes I and II) which also demonstrates the propensity of the 2-Me₂N to be axial and shows the free energy difference between chair and twist conformations to be very small.

We assign the axial preference of the Me₂N in 13 and 14 in part to repulsive interactions between equatorial Me₂N and the phenyl substituent at N(3) in conformation 21, as proposed by Nifantiev⁴ for 4. This effect is in direct contrast to the repulsions assigned previously to these two substituents in their four-coordinate, 2-oxo-1,3,2oxazaphosphorinane counterparts in the conformation analogous to 20 in which the Me₂N is axial.^{2a-e} These findings demonstrate the variety of steric and stereoelectronic interactions operative in the conformational equilibria of 1,3,2-oxazaphosphorinanes and the surprising differences in the conformational properties of this ring system when it contains three- rather than four-coordinate phosphorus.

Results

Preparation. 1,3,2-Oxazaphosphorinanes 7-10 resulted in 81-91% yields from the reaction of the appropriate





^{(2) (}a) Bajwa, G. S.; Bentrude, W. G.; Pantaleo, N. S.; Newton, M. G.; Hargis, J. H. J. Am. Chem. Soc. 1979, 101, 1602-1604. (b) Chandrasekaran, S.; Bentrude, W. G. Tetrahedron Lett. 1980, 21, 4671-4674. (c) Bajwa, G. S.; Chandrasekaran, S.; Hargis, J. H.; Sopchik, A. E.; Blatter, D.; Bentrude, W. G. J. Am. Chem. Soc. 1982, 104, 6385-6392. (d) Bentrude, W. G.; Day, R. O.; Holmes, J. M.; Quin, G. S.; Setzer, W. N.; Sopchik, A. E.; Holmes, R. R. J. Am. Chem. Soc. 1984, 106, 106-111. (e) Holmes, R. R.; Day, R. O.; Setzer, W. N.; Sopchik, A. E.; Bentrude, W. G. J. Am. Chem. Soc. 1984, 106, 2353-2358. (f) Setzer, W. N.; Sopchik, A. E.; Bentrude, W. G. J. Am. Chem. Soc. 1985, 107, 2083-2091. (g) Bentrude, W. G.; Setzer, W. N.; Sopchik, A. E.; Bajwa, G. S.; Burright, D. D.; Hutchinson, J. P. J. Am. Chem. Soc. 1986, 108, 6669-6675. (h) Bentrude, W. G.; Setzer, W. N.; Sopchik, A. E.; Chandrasekaran, S.; Ashby, M. T. J. Am. Chem. Soc. 1988, 107, 7119-7127. (i) Bentrude, W. G.; Setzer, W. N.; Newton, M. G.; Meehan, E. J.; Ramli, E.; Khan, M.; Ealick, S. Phosphorus, Sulfur, Silicon 1991, 57, 25-35. (j) Bentrude, W. G.; Setzer, W. N.; Newton, M. G.; Meehan, E. J.; Ramli, E.; Khan, M.; Ealick, S. Phosphorus, Sulfur, Silicon 1991, 57, 25-35. (j) Bentrude, W. G.; Setzer, W. N.; Kergaye, A. A.; Ethridge, V.; Saadein, M. R.; Arif, A. M. Phosphorus, Sulfur, Silicon 1991, 57, 37-49. (k) Bentrude, W. G.; Setzer, W. N.; Khan, M.; Sopchik, A. E.; Ramli, E. J. Org. Chem. 1991, 56, 6127-6131. (a) See, for example: Nelson, K. A.; Bentrude, W. G.; Setzer, W. N.;

⁽³⁾ See, for example: Nelson, K. A.; Bentrude, W. G.; Setzer, W. N.; Hutchinson, J. P. J. Am. Chem. Soc. 1987, 109, 4058-4064 and references cited therein.

^{(4) (}a) Nifant'ev, E. E.; Borisenko, A. B.; Sorokina, S. F.; Grachev, M. K.; Zavalishina, A. I. Zh. Obshch. Khim. 1977, 47, 2474-2480 and references cited therein. (b) Milliaresi, E. E.; Kharshan, M. A.; Preobrazhenskaya, E. A.; Nifant'ev, E. E. Zh. Obshch. Khim. 1984, 54, 1049-1051 and references cited therein.

⁽⁵⁾ Majoral, J.-P.; Bergounhou, C.; Navech, J. Bull. Chim. Soc. Fr. 1973, 3146-3149.

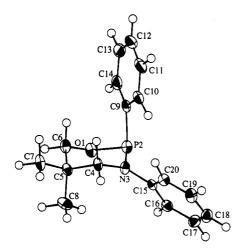


Figure 1. ORTEP plot for 11.

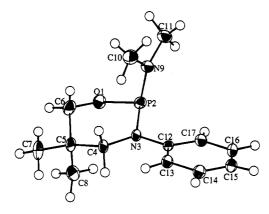
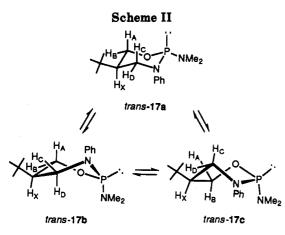


Figure 2. ORTEP plot for 13.



2-chloro-3-phenyl-1,3,2-oxazaphosphorinane with ROH in the presence of Et₃N. Compound 11 was prepared by the reaction of bis(N,N-diethylamino) phenylphosphine with the amino alcohol in 67% yield. Compounds 13-16 were synthesized in 47-73% yield by the reaction of the amino alcohol with hexamethylphosphorous triamide. The reaction of the 2-chloro-1,3,2-oxazaphosphorinane with methylamine and dimethylamine afforded 12 and 17. Compound 17 was formed as a mixture of cis and trans diastereomers. The ¹H NMR parameters of 17 were obtained from spectra on materials with cis/trans ratios of 18/82 (freshly made) which changed to 64/36 on distillation. The latter mixture was further equilibrated on standing in $C_6 D_6$ at room temperature to an 80/20 (cis/ trans) ratio (taken to be the equilibrium ratio) for determination of the parameters for cis-17.

Table I. Crystal Data for 11 and 13 at -125 °C

Table 1. Crystal Data for 11 and 15 at -125 C							
compd	11	13					
mol formula	C ₁₇ H ₂₀ NOP	C ₁₃ H ₂₁ N ₂ OP					
mol wt	285.329	252.299					
space grp	$P2_1/n$ (No. 14)	$P2_1/n$ (No. 14)					
cryst system	monoclinic	monoclinic					
cell dimens							
a, Å	15.207(1)	13.581(2)					
b, Å	5.942(1)	6.024(1)					
c. A	18.098(1)	16.315(2)					
α , deg							
β , deg	113.35(1)	90.70(1)					
γ , deg							
ν, Å ³	1501.40	1334.73					
Ż	4.0	4.0					
$D_{\rm calcd}, {\rm g/cm^3}$	1.262	1.255					
radiation, Å	λ (Cu) 1.54180	λ (Cu) 1.54180					
2θ range, deg	4.00-130.00	4.00-130.00					
scan technique	$\theta/2\theta$	$\theta/2\theta$					
scan width, deg	0.8000 +	0.8000 +					
	$0.1400 \tan \theta$	$0.1400 \tan \theta$					
no. of refins used	2128	2505					
absorption coeff, cm ⁻¹	15.659	17.105					
data to param ratio	9.195	9.166					
shift to error ratio	0.000	0.004					
R	0.0453	0.0498					
R _w	0.0495	0.0547					

Table II. Selected Bond Lengths (Å), Bond Angles (deg), and Torsion Angles (deg) for 11 and 13

	compd				
atoms	11	13			
P(2)-O(1)	1.633(2)	1.631(2)			
P(2)-N(3)	1.689(2)	1.727(3)			
P(2)-N(9)		1.667(3)			
P(2)-C(9)	1.844(3)				
O(1)-C(6)	1.451(3)	1.441(5)			
N(3)-C(4)	1.474(3)	1.464(4)			
O(1)-P(2)-N(3)	101.1(1)	99.3(1)			
P(2)-O(1)-C(6)	118.8(2)	123.4(2)			
P(2)-N(3)-C(4)	119.9(2)	125.8(2)			
P(2)-N(3)-C(12)		116.4(2)			
C(4)-N(3)-C(12)		115.7(3)			
P(2)-N(3)-C(15)	119.4(2)				
C(4)-N(3)-C(15)	119.7(2)				
P(2)-N(9)-C(10)		127.2(2)			
P(2)-N(9)-C(11)		116.5(3)			
C(10)-N(9)-C(11)		114.0(3)			
O(1)-C(6)-C(5)	112.5(2)	110.5(3)			
N(3)-C(4)-C(5)	110.6(2)	113.6(3)			
C(4)-C(5)-C(6)	109.1(2)	108.2(3)			
P(2)-O(1)-C(6)-H(3)	-63.01(2.53)	-52.38(2.90)			
P(2)-O(1)-C(6)-H(4)	179.24(2.27)	174.70(3.10)			
P(2)-N(3)-C(4)-H(1)	65.95(2.56)	92.18(2.78)			
P(2)-N(3)-C(4)-H(2)	-175.72(2.64)	-153.43(3.17)			
N(3)-P(2)-N(9)-C(10)		40.41(0.30)			
N(3)-P(2)-N(9)-C(11)		-158.26(0.22)			
O(1)-P(2)-N(9)-C(10)		-63.64(0.30)			
O(1)-P(2)-N(9)-C(11)		97.68(0.24)			

Single-Crystal X-ray Structures of 11 and 13. ORTEP perspective views of 11 and 13 are given in Figures 1 and 2. Crystal data for 11 and 13 are compiled in Table I. Selected bond distances, bond angles, and torsion angles are shown in Table II.

The ORTEP structures of 11 and 13 show that they are in the chair conformation in the solid state. The sixmembered ring of 13 is somewhat more distorted from a perfect chair geometry (see below) than is that of 11. The sums of the bond angles around N(3) and N(9) of 13 are 357.9° and 357.7°, respectively. This indicates that the nitrogen atoms approach sp² hybridization. (The sum of the bond angles for approximately sp³-hybridized NH₃ is 321.9°.⁷) However, one of the bond angles about N(3) and N(9) is larger than 120° while the others are less ($\angle P$ -(2)N(3)C(4) = 125.8 (2)°, $\angle P$ (2)N(3)C(12) = 116.4 (2)°,

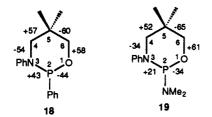
Table III. Selected ¹H NMR Coupling Constants (Hz) for 7-17^a

compd	solvent	⁸ J _{AP}	⁸ J _{BP}	³ J _{CP}	³ J _{DP}	⁴ J _{BD}	³ J _{AX}	³ J _{BX}	³ J _{CX}	³ J _{DX}	³ J _{AY}	⁸ J _{BY}	⁸ J _{CY}	⁸ J _{DY}
·	C ₆ D ₆	7.7	9.2	3.7	<u>ь</u>									
4					-	0.9	9.9	4.4	10.1	4.4	3.3	4.6	3.7	4.7
4	CD ₃ CN	7.2	9.3	3.8	5.2	-0.8	10.3	4.5	10.3	4.5	3.3	4.3	3.5	4.4
7	C_6D_6	3.7	12.6	1.8	5.5	-2.1								
7	CD_3CN	3.1	12.6	1.1	5.4	-2.2								
8	C_6D_6	3.7	13.5	2.9	6.4	-2.2								
8	CD ₃ CN	3.8	13.2	2.9	6.3	-2.2								
9¢	$C_6 D_6$	3.5	11.9	1.8	5.2	-1.5	12.0	4.4	12.0	4.5	2.8	2.8	2.9	3.3
10°	C ⁶ D ₆	3.6	12.4	2.9	5.9	-1.4	11.5	4.5	11.6	4.4	2.9	3.1	2.9	4.3
11 ^d	$C_6 D_6$	3.4	11.8	<0.5	2.9	-2.2								
11	CD ₃ CN	3.4	11.8	<0.5	3.3	-2.1								
12	$C_{6}D_{6}$	4.9	12.0	1.3	5.3	-2.1								
13	C_6D_6	7.4	10.6	2.1	4.6	-1.8								
13	CD ₃ CN	7.0	10.8	1.5	4.9	-1.8								
14°	$C_6 D_6$	8.0	9.7	1.8	4.5	-0.9	9.8	4.8	10.7	4.6	3.7	4.6	3.6	4.5
15 ^d	$C_6 D_6$	8.7	10.5	2.3	4.3	-1.6								
15	CD ₃ CN	7.7	10.8	1.4	4.7	-1.8								
16 ^d	C ₆ D ₆	8.6	10.3	3.9	4.5	-1.5								
16	CD ₃ CN	7.7	10.5	3.3	5.0	-1.8								
cis-17 ^e	C ₆ D ₆	11.9	8.9	1.2	5.0	-1.3	8.7	5.7	11.4	4.8				
trans-17e	C_6D_6	7.0	12. 9	2.5	7.2	-0.7	9.0	6.6	9.3	5.2				

^a At 300 MHz, ambient temperature, unless otherwise noted. ^b Overlapped with Me₂N signal. ^c Iteratively refined with LAOCN5 program: rms error, 0.136 Hz (9), 0.099 Hz (10), 0.072 Hz (14); probable error, 0.012–0.018 Hz (9), 0.009 Hz (10), 0.006–0.010 Hz (14); J 0.017–0.039 Hz (9), 0.013–0.019 Hz (10), 0.009–0.017 Hz (10). ^d Simulated by use of LAOCN5. ^e At 500 MHz.

 $\angle C(4)N(3)C(12) = 115.7 (3)^{\circ}; \angle P(2)N(9)C(10) = 127.2 (2)^{\circ}, \angle P(2)N(9)C(11) = 116.5 (3)^{\circ}, \angle C(10)N(9)C(11) = 114.0 (3)^{\circ}$). Further distortion is seen in torsion angles P(2)O-(1)C(6)H_A and P(2)O(1)C(6)H_B, -52.4° and 174.7°, respectively. On the other hand, the sum of the bond angles around N(3) of 11 is 359.0°, and each bond angle is almost the same indicating that the nitrogen atom is truly sp² hybridized and that the six-membered ring of 11 is less distorted than that of 13. For 11, the torsion angles of P(2)O(1)C(6)H_A and P(2)O(1)C(6)H_B are -63.0° and 179.2°, respectively, close to those for an idealized, undistorted, cyclohexane geometry, -60° and 180°, respectively.

The ring torsion angles for 11 and 13 are given in structures 18 and 19, respectively. Angles O(1)P(2)N(3)C(4) and P(2)N(3)C(4)C(5) of 13 are about 20° less than those of 11. The N(3)P(2)O(1)C(6) angle of 13 is reduced by 10°. This indicates that the distortion of the ring of 13 is best characterized as *flattening at the* N(3)-P(2) end.

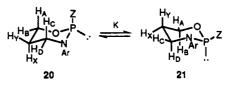


¹H NMR Parameters and Conformations of 7-16. In Tables III and IV are listed the pertinent ¹H NMR coupling constants and chemical shifts for 7-16. Protons on the six-membered ring are designated H_A , H_B , H_C , H_D , H_X , and H_Y , as shown in 20 and 21. For 5,5-dimethyl compounds 7, 8, and 11-13, 15, and 16, H_X and H_Y are replaced by methyl groups. As the values of ${}^{3}J_{PNCH}$ have not been found to be useful in conformational analyses of three-coordinate phosphorus containing heterocyclic sixmembered rings,⁸ only the values of ${}^{3}J_{POCH}$ and ${}^{3}J_{HH}$ are used to determine the conformations of these six-membered rings.

Table IV. Selected ¹H NMR Chemical Shifts (ppm) for

7-17*							
compd	solvent	Α	В	C	D	х	Y
4	C ₆ D ₆	4.07	3. 6 3	3.04	2.52	1.60	1.23
4	CD ₃ CN	4.09	3.65	3.21	2.83	1.78	1.56
7	C_6D_6	3.98	3.18	3.50	2.66		
7	CD ₃ CN	4.00	3.33	3.55	2.92		
8	C_6D_6	3.97	3.17	3.41	2.48		
8	CD ₃ CN	4.05	3.50	3.62	2.96		
9	C_6D_6	4.07	3.50	3.57	2.88	1.90	1.15
10	C_6D_6	4.00	3.45	3.38	2.69	1.68	1.08
11	C_6D_6	3.61	3.28	3.19	3.06		
11	CD ₃ CN	3.62	3.45	3.28	3.38		
12	$C_6 D_6$	3.64	3.16	2.98	2.80		
13	C_6D_6	3.80	3.25	3.29	2.92		
13	CD ₃ CN	3.87	3.35	3.48	3.15		
14	C_6D_6	3.87	3.56	3.39	3.12	1.69	1.24
15	C_6D_6	3.65	3.22	3.12	2.82		
15	CD ₃ CN	3.87	3.41	3.47	3.21		
16	C_6D_6	3.85	3.36	3.38	2.98		
16	CD ₃ CN	3.81	3.35	3.43	3.01		
cis-17 ^b	C_6D_6	3.90	3.77	3.52	3.44	1.72	
trans-17 ^b	C_6D_6	3.73	3. 9 6	3.26	3.50	1.93	

 a At 300 MHz, ambient temperature, unless otherwise noted. b At 500 MHz.



The small ${}^{3}J_{AP}$ (3.1-3.8 Hz) and large ${}^{3}J_{BP}$ (11.8-13.5 Hz) recorded for 7-11 (Table III) are similar to those seen for the corresponding three-coordinate 1,3,2-dioxaphosphorinanes with Z = MeO, (CF₃)₂CHO, and Ph¹ and allow 7-11 to be assigned to a single chair conformation (20). In addition, the large ${}^{3}J_{AX}$ and ${}^{3}J_{CX}$ (11.5, 12.0 Hz for diaxial protons) and small ${}^{3}J_{BX}$ and ${}^{3}J_{DX}$ (4.4, 4.5 Hz for gauche protons) for 9 and 10 are typical values for vicinal couplings of chair-form rings. Furthermore, relatively large long-range W-configuration couplings, ${}^{4}J_{BD}$ (-1.4 to -2.2 Hz), are observed for 7-11, as expected for 20. 1,3,2-Oxaza-phosphorinane 12 also largely populates a single chair with the NHMe group axial on phosphorus, as evidenced by

⁽⁷⁾ Wade, L. G. Organic Chemistry, Prentice-Hall, Inc.: New York, 1987; pp 55.

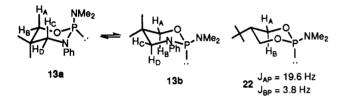
⁽⁸⁾ Hutchins, R. O.; Maryanoff, B. E.; Albrand, J. P.; Cogne, A.; Gagnaire, D.; Robert, J. B. J. Am. Chem. Soc. 1972, 94, 9151-9158.

Equilibria of 2-Z-3-Phenyl-1,3,2-oxazaphosphorinanes

 ${}^{3}J_{AP}$ (4.9 Hz) and ${}^{3}J_{BP}$ (12.0 Hz). However, for 13–16 ${}^{3}J_{AP}$ (7.0–8.7 Hz) is increased by about 4 Hz compared to ${}^{3}J_{AP}$ for 7–11. Likewise, ${}^{3}J_{BP}$ (9.7–10.8 Hz) is decreased by approximately 2 Hz. The values of ${}^{3}J_{AX}$ and ${}^{3}J_{CX}$ for 14 (9.8 and 10.7 Hz, respectively) also are reduced and ${}^{3}J_{BY}$ (4.6 Hz) and J_{DY} (4.5 Hz) are appropriately increased. Thus, although 20 remains the major conformer populated, both conformers of the chair-chair equilibrium 20 \Rightarrow 21 are present in solution for 13-16 (Z = Me₂N). Clearly, the values of ${}^{3}J_{CP}$ and ${}^{3}J_{DP}$ show no trends useful in discerning the population of 20 vs 21.

For 15 and 16, two different substituents are attached to the N-phenyl group, the electron-withdrawing m-CF₃, and the electron-donating p-NMe₂. However, there is a near- constancy of the ${}^{3}J_{AP}$ and ${}^{3}J_{BP}$ values for both compounds, which indicates a lack of either an electronic substituent or solvent effect on the equilibrium 20 \approx 21 (Table III).

Chair-Chair Equilibrium Constants for 13-16. The equilibrium constants $(K, 20 \Rightarrow 21)$ can be estimated for 13-16 on the basis of the observed coupling constants ${}^{3}J_{AP}$ and ${}^{3}J_{BP}$. The mole fraction of 13a (N(13a)) can be readily estimated.



Thus, it is clear that

1

$$N(13a)^{3}J_{AP}(13a) + N(13b)^{3}J_{AP}(13b) = {}^{3}J_{AP}(\text{obsd})$$
 (1)

$$N(13a) = 1 - N(13b)$$
 (2)

therefore,

$$N(13a) = ({}^{3}J_{AP}(\text{obsd}) - {}^{3}J_{AP}(13b)) / ({}^{3}J_{AP}(13a) - {}^{3}J_{AP}(13b))$$
(3)

Similarly, for ${}^{3}J_{\rm BP}$

$$N(13a) = ({}^{3}J_{\rm BP}({\rm obsd}) - {}^{3}J_{\rm BP}(13b))/({}^{3}J_{\rm BP}(13a) - {}^{3}J_{\rm BP}(13b))$$
(4)

The values of ${}^{3}J_{AP}(13b)$ and ${}^{3}J_{BP}(13b)$ in eqs 3 and 4 are for the specific conformation 13b and can be assumed to be those for 22⁹ (19.6 and 3.8 Hz, respectively), which is known to be in the chair conformation shown. (The axial lone-pair orientation greatly enhances ${}^{3}J_{AP}$). However, ${}^{3}J_{AP}(13a)$ and ${}^{3}J_{BP}(13a)$ for conformation 13a cannot be assumed from model compounds because of the ring distortion present in 13a (Figure 2). However, they could be calculated from Karplus eq 5 were the constants A and B known.

$${}^{3}J_{\rm POCH} = A\,\cos^2\phi_{\rm HP} + B\cos\phi_{\rm HP} \tag{5}$$

To determine A and B, we chose 11 as the model compound. As noted above, the six-membered ring of 11 is relatively undistorted in the solid phase (Figure 1) with the phenyl group attached axially to phosphorus atom. It

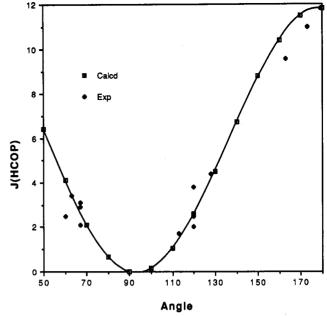


Figure 3. Dihedral angle dependence of ${}^{3}J_{HP}$ for three-coordinate 1,3,2-dioxaphosphorinanes. \Box are points calculated from eq 5 with A = 13.3, B = 1.47. \blacklozenge are experimental data from ref 10.

can be assumed, based on its $J_{\rm HP}$ values, that 11 in solution is essentially 100% in the same chair conformation. Thus, constants A and B in eq 5 can be determined from the observed ${}^{3}J_{AP}$ and ${}^{3}J_{BP}$ values of 11 (3.4 and 11.8 Hz, respectively) and the measured torsion angles, $\phi_{AP} = 63^{\circ}$, $\phi_{BP} = 179^{\circ}$, from the crystal structure of 11. They were found to be 13.3 (A) and 1.47 (B). The reliability of this approach was checked by plotting ${}^{3}J_{POCH}$ vs dihedral angle (Figure 3) from the Karplus curve (□) generated from eq 5 (A = 13.3 and B = 1.47) along with experimental data from the literature (•).¹⁰ There is a reasonably good agreement between the curve calculated by eq 5 (50° \leq $\phi_{\rm PH} \leq 180^{\circ}$) and the limited number of experimental values available. This equation assumes ${}^{3}J_{\rm HP}$ is zero at 90° which may not be true. However, near the angles measured for 13 it is considered to be adequate, since the experimental points fall reasonably near the curve in the critical area of about 60-70°, just above the 52° angle for ϕ_{AP} of 13. The value for $J_{\rm BP}$ of 13a in fact could have been assumed from that for 11 (11.8 Hz), since the angles ϕ_{AP} for 11 and 13a are very close (175°, 179°), and a Karplus curve will not be steep in the region 175-180°.

The values of ${}^{3}J_{AP}$ (5.8 Hz) and ${}^{3}J_{BP}$ (11.7 Hz) for 13a were calculated from eq 5 and torsion angles ϕ_{AP} (52°) and ϕ_{BP} (175°) taken from the crystal structure of 13. (Note that the major effect of ring distortion is on ${}^{3}J_{AP}$.) The mole fraction of 13a was calculated to be 88% from eq 3 (${}^{3}J_{AP}$) and 86% from eq 4 (${}^{3}J_{BP}$), in excellent internal agreement. Similarly, for 14–16 the chair-chair equilibrium constants (K) were calculated on the assumption that the six-membered rings of conformer 20 are distorted to the same degree as those for 13a. The assumed coupling constants were as follows: ${}^{3}J_{AP}(20) = 5.8$ Hz, ${}^{3}J_{BP}(20) = 11.7$ Hz, ${}^{3}J_{AP}(21) = 19.6$ Hz, ${}^{3}J_{BP}(21) = 3.8$ Hz. The calculated results are recorded in Table V.

Chair-Twist Equilibria for cis- and trans-17. ¹H NMR coupling constants for cis- and trans-17 are quite different from those for 7-16 (Table III). The values of

⁽⁹⁾ Bentrude, W. G.; Tan, H.-W. J. Am. Chem. Soc. 1973, 95, 4666–4675. Values of $J_{\rm HP}$ of 19.5–21.5 Hz have been found for $J_{\rm AP}$ of the t-BuP analog of 22 (ref 13a) and for other three-coordinate 1,3,2-oxazaphos-phorinanes with R₂N on phosphorus that are clearly in a single chair conformation like 13b (Huang, Y., Mullah, N. Unpublished results).

⁽¹⁰⁾ White, D. W.; Verkade, J. G. J. Magn. Reson. 1970, 3, 111-116 and references cited therein.

Table V. Estimated Equilibrium Constants (K = 21/20) at 25 °C

		20 on obsda			∆G°	
compd	$J_{\rm AP}$	$J_{ m BP}$	avg	K	(kcal/mol)	
13	88	86	87	0.15	1.1	
14	84	75	80	0.25	0.82	
15	7 9	85	82	0.22	0.90	
16	80	82	81	0.23	0.87	
4 ^b	86	68	77	0.30	0.71	
26°	31	28	30	2.3	-0.50	

^a See text for assumed ${}^{3}J_{AP}$ and ${}^{3}J_{BP}$ for 20 and 21. ^b Based on coupling constants measured at 300 MHz in C₆D₆. ^c Reference 16.

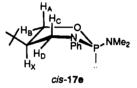
 ${}^{3}J_{\rm HP}$ and ${}^{3}J_{\rm HH}$ show that *cis*- and *trans*-17 populate nonchair conformations, at least in part. A similar approach, using assumed ${}^{3}J_{\rm HP}$ or ${}^{3}J_{\rm HH}$ values for various conformers, allows estimates of conformer populations for *cis*-17 and *trans*-17 to be made.

For the nonchair (twist/boat) forms depicted in Schemes I and II, one side of the ring remains in a geometry about the C4–C5 or C5–C6 bond that is the same as the chair conformation from which it is formed. The protons attached to C4 or C6 remain axial or equatorial as in the chair. They also move into positions synclinal and antiperiplanar, respectively, with respect to phosphorus and become fully pseudoaxial and pseudoequatorial if the degree of twisting of the ring moves the conformation fully into a boat form on the boat-twist-boat pseudorotational cycle. These geometrical considerations apply, for example, to the oxygen side $(H_A \text{ and } H_B)$ of the ring in *cis*-17c and trans-17b and the nitrogen side $(H_C \text{ and } H_D)$ of cis-17d and trans-17c. At the same time the methylene protons on the opposite side of the ring move from axial and equatorial toward pseudoequatorial and pseudoaxial positions, respectively. Simultaneously, the lone pair on phosphorus moves from equatorial in cis-17a toward the pseudoaxial position in its twist/boat forms and vice versa for the lone pair of trans-17a. The degree of twist, for example, in cis-17c and cis-17d, determines the extent to which the methylene protons and lone pair fully assume their new positions. In true boat conformations, certain of the H-C-C-H and H-C-O-P torsion angles are of the order 60° or 180° as in a chair, and values of ${}^{\bar{3}}J_{\rm HP}$ and ${}^{3}J_{\rm HH}$ can be assumed from three-coordinate 1,3,2-oxazaphosphorinanes (Table III) or 1,3,2-dioxaphosphorinanes that populate a single chair conformation.

From the inspection of the observed, time-averaged values of ${}^{3}J_{HH}$ and ${}^{3}J_{HP}$ for cis-17 and trans-17 (Table III), it is relatively easy to discern qualitatively the nature of the conformational equilibria involved. Thus, for cis-17 it is clear that cis-17c and cis-17b are not populated or the value of ${}^{3}J_{CX}$ would be reduced from the 11.4 Hz value observed which is typical of trans diaxial protons in chair forms similar to cis-17a (Table III). However, in cis-17d H_{C} and H_{X} remain antiperiplanar and essentially trans diaxial. The increase in ${}^{3}J_{AP}$ observed to a time-averaged value (11.9 Hz), much larger than the 5.8 Hz value predicted for cis-17a by the above Karplus curve analysis of such rings with axial Me₂N, shows unmistakably that cis-17d is populated to a significant degree. This also leads to a reduction in ${}^{3}J_{BP}$ (8.9 Hz) from the close to 12 Hz value found in Table III for similar equatorial hydrogens on chair-form rings with axial substituents on phosphorus (7-12) and from the 11.7 Hz value estimated for 13 from the above Karplus curve considerations and presumably applicable to cis-17a.

For trans-17 there are only three possible conformers in equilibrium worthy of consideration (Scheme II) because the diaxial-substituted chair form would be too high in energy. The observed reductions in ${}^{3}J_{AX}$ and ${}^{3}J_{CX}$ mean that trans-17a is depopulated. The near-equal values of ${}^{3}J_{AX}$ (9.0 Hz) and ${}^{3}J_{CX}$ (9.3 Hz) indicate that trans-17b and trans-17c are approximately equally populated, if the degree of twist is about equal in each; i.e., ${}^{3}J_{CX}$ for trans-17b is equal to ${}^{3}J_{AX}$ for trans-17c. The accompanying increase in ${}^{3}J_{BX}$ (6.6 Hz) from the 4-5 Hz value for an equatorial hydrogen of the chair form ring of 9, 10, or 14 (Table III) is fully expected if the ring of trans-17c is twisted far enough for H_B and H_X to become nearly eclipsed, and ${}^{3}J_{BX}$ for trans-17c is therefore large. The increase in ${}^{3}J_{DX}$, though smaller, is similarly explained by trans-17b.

Estimation of actual conformer populations for the above equilibria must be done with caution and care, because the coupling constants to hydrogen and phosphorus for pseudoaxial and pseudoequatorial protons depend in a Karplus equation dependent fashion on the degree of twisting, as noted above. It appears from Dreiding models that the estimation of conformer populations from measured ${}^{3}J_{\rm HP}$ values is likely to be more accurate than if observed ${}^{3}J_{\rm HP}$ values are used. This results first from the greater validity of assumed ${}^{3}J_{\rm HP}$ values. Thus, a rotation of about 60° about the C–O and C–N bonds of *cis*-17e is required to convert it to one of the boat



conformations (cis-17c or cis-17d) on either side of it on the pseudorotational pathway. In so doing the protons at C4 and C6 assume fully pseudoaxial or pseudoequatorial positions for which assumed ${}^{3}J_{\rm HP}$ values can be taken from those of axial and equatorial protons of model chair-form molecules. Further rotation does not change the HCOP (the angle of prime interest) or HCNP torsion angle. By contrast the HCCH torsion angles related to C4-C5 and C5-C6 bond rotation continue to change on the way to the next true boat conformation (beyond cis-17c or cis-17d). Secondly, the differences between ${}^{3}J_{HP}(obsd)$ and the values expected for the chair forms cis-17a and trans-17a (i.e., the changes in ${}^{3}J_{HP}$ (obsd)) are considerably greater than those seen when considering ${}^{3}J_{\rm HH}$ values (Table III) which leads to more accurate calculations. (As for 7-16 variations in ${}^{3}J_{CP}$ and ${}^{3}J_{DP}$ show no useful trends). Therefore, in the estimates of equilibria made below, we rely first on the measured ${}^{3}J_{\rm HP}$ values and then check those numbers against those obtained from observed ${}^{3}J_{\rm HH}$.

For the equilibrium cis-17a \Rightarrow cis-17d, one may reasonably assume the values for cis-17a estimated by the Karplus relation for 13 (rounded off to the nearest Hz: ${}^{3}J_{AP} = 6$ Hz; ${}^{3}J_{BP} = 12$ Hz). For cis-17d those for 22 (similarly rounded off) are reasonable if cis-17d is sufficiently twisted so that H_A and H_B are essentially pseudoequatorial and pseudoaxial, respectively: ${}^{3}J_{AP} = 20$ Hz; ${}^{3}J_{BP} = 4$ Hz. It is readily shown, by the algebra used to obtain eqs 3 and 4, that

$$N(17a) = ({}^{3}J_{AP}(\text{obsd}) - {}^{3}J_{AP}(17d)) / ({}^{3}J_{AP}(17a) -$$

 ${}^{3}J_{\rm AP}(17{\rm d}))$ (6)

where the terms are as defined for eqs 1-4 and refer only

to the conformers of Scheme I representing cis-17. A similar equation can be written for ${}^{3}J_{BP}$. From eq 6 and its ${}^{3}J_{\rm BP}$ equivalent, N(17a) values of 0.58 and 0.61 are calculated based on observed ${}^{3}J_{AP}$ and ${}^{3}J_{BP}$, respectively. This is very good agreement and says that about 60% of cis-17 remains in the chair form, cis-17a, with the remaining 40% in boat/twist form 17d. If ${}^{3}J_{AX}$ for cis-17a is 11.5 Hz, as in 22, and has a value of 4.5 Hz in cis-17d, ${}^{3}J_{AX}(obsd)$ is calculated for a 60/40 ratio of cis-17a/cis-17d to be 8.7 Hz, the experimentally observed value. We have no good model compound from which to assume ${}^{3}J_{AX}$ for cis-17d, as its H_A-C6-C5-H_X torsion angle is approximately 120° if *cis*-17**d** is a boat form as required for the assumed values of ${}^{3}J_{\rm HP}$ to be reasonably correct. However, even if the value of ${}^{3}J_{AX}$ for *cis*-17d is only 3.7 Hz, the calculated ${}^{3}J_{AX}$ would still be 8.4 Hz, nearly within the experimental error of the observed 8.7 Hz value.

The three-conformer equilibrium of Scheme II can be treated rather simply algebraically if it is assumed that N(trans-17b) = N(trans-17c) as proposed above. This means that

$$N(trans-17a) + (2N(trans-17b)) = 1$$

Since for J_{AP}

 $N(17a)J_{AP}(17a) + N(17b)J_{AP}(17b) +$ $N(17c)J_{AP}(17c) = J_{AP}(obsd)$

it is easy to derive expression 7 for trans-17 under conditions that N(17b) = N(17c):

$$N(17b) = (J_{AP}(17a) - J_{AP}(obsd))/(2J_{AP}(17a) - J_{AP}(17b) - J_{AP}(17c))$$
(7)

Assuming that trans-17b and trans-17c are twisted such that H_A and H_B are truly pseudoequatorial and pseudoaxial with respect to phosphorus in trans-17b and vice versa for trans-17c, then reasonable values for ${}^{3}J_{AP}$ (rounded off to the nearest Hz) can be assumed from those of 22 and the rings of Table III that are completely in chair conformations (i.e., 7-12): $J_{AP}(17a) = 4 \text{ Hz}; J_{AP}(17b) = 4 \text{ Hz}; J_{AP}(1$ (17c) = 12 Hz. These values lead from eq 7 to N(trans-(17b) = N(trans-17c) = 0.38. The analogous expression for J_{BP} and assumed values: $J_{BP}(17a) = 20$ Hz; $J_{BP}(17b)$ = 12 Hz; $J_{BP}(17c)$ = 4 Hz gives N(trans-17b) = N(trans-17b)17c) = 0.30. The average of these two estimates is 0.34. Thus, one can say that a roughly equal amount of each conformer of Scheme II is populated by trans-17. As a cross check, use of the reasonable values for ${}^{3}J_{AX}$ for the 17a and 17b conformers of trans-17 of 12 Hz, along with the 4.5 Hz ${}^{3}J_{AX}$ value assumed for *cis*-17d, predicts ${}^{3}J_{AX}$ for trans-17 to be 9.5 Hz, compared to the 9.0 Hz experimental value.

One must again state that the estimates made above are only approximate because of the errors in assumed couplings. However, the agreements reached using J_{AX} along with J_{AP} and J_{BP} are gratifying. Also, as outlined in the Discussion, the equilibria of Schemes I and II can be reasonably well interpreted in terms of the steric effects on the chair-chair equilibria observed for PhN(3)/Me₂N compounds (Table III). For these interpretations, the assignment of conformer populations need not be precise.

³¹P Chemical Shifts for 7-17. The ³¹P NMR chemical shifts of 7-17 are tabulated in Table VI. Individual signals were observed for each diastereomer of 17. The upfield signal was assigned to the cis diastereomer.^{2c} This assignment was confirmed by stereospecific retentive

Table VI. ³¹P NMR Chemical Shifts (ppm) for 7-17 in C.D.ª

- 0 0							
δ	compd	δ	compd	δ			
125.5	11	111.9	15	120.8			
131.1	12	108.6	16	124.3			
133.7	13	120.7	cis-7	124.2			
138.7	14	126.9	trans-17	127.0			
	125.5 131.1 133.7	δ compd 125.5 11 131.1 12 133.7 13	δ compd δ 125.5 11 111.9 131.1 12 108.6 133.7 13 120.7	δ compd δ compd 125.5 11 111.9 15 131.1 12 108.6 16 133.7 13 120.7 cis-7			

^a Positive chemical shifts are downfield from the external 85% H₃PO₄. At 121 MHz, ambient temperature.

oxidation of 17 to the respective cis and trans isomers of 2-oxo-2-(dimethylamino)-3-phenyl-5-tert-butyl-1,3,2-oxazaphosphorinane whose identities were established previously.^{2c} The so-called δ substituent effect¹¹ in ³¹P NMR spectroscopy is observed for 7-14. This effect arises from the presence of the equatorial methyl of the gem dimethyl substitution at C(5) and the axial substituent on three-coordinate phosphorus and results in an approximately 7.5 ppm upfield chemical shift in three-coordinate 1,3,2-dioxaphosphorinanes.¹¹ Thus, for 7, 8, and 13, the gem dimethyl substitution at C(5) is accompanied by a 6.2-8.2 ppm upfield ³¹P shift compared to unsubstituted 9, 10, and 14. This is consistent with the postulated predominant axial orientation of the Z substitutent of 7-14. For cis- and trans-17, the combination of δ substituent effects (5.5 ppm upfield for the equatorial 5-tert-butyl substituent¹¹) and cis/trans effects (cis, P-Z axial upfield of trans, P-Z equatorial^{1b}) should generate a large difference in chemical shift if they were both in chair conformations. However, only a 2.8 ppm difference between cis- and trans-17 was observed. This is further evidence for the chair-twist equilibria proposed for cisand trans-17 in which the Me₂N is moved out of the axial position in cis-17d but becomes pseudoaxial in trans-17b and trans-17c.

Discussion

The position of the equilibrium $20 \Rightarrow 21$ and, therefore. the orientation of a substituent (Z) on phosphorus are determined by a balance of steric interactions and sterecelectronic factors.¹² The three potentially relevant steric factors are as follows: repulsive 1,3-syn axial (Z/H) interactions (destabilizing conformer 20); repulsions between vicinal diequatorial substituents on N(3) and P(e.g., Ph/Z_{eq}) destabilizing conformer 21; and repulsions between vicinal axial-equatorial (Ph/Zax) substituents (destabilizing conformer 20). Stereoelectronic factors of potential consequence include stabilizing $n-\sigma^*$ interactions of the type likely involved in the endo anomeric effect.¹² Thus, an axial P-Z orientation (20) would be stabilized by oxygen (n_0/σ^*_{PZ}) and nitrogen (n_N/σ^*_{PZ}) lone pair overlap with the antibonding P-Z orbital (structure 23). This stabilization might be further aided by interaction of the equatorial lone pair on phosphorus with endocyclic C-O and C-N antibonding orbitals, a factor not always considered. Back donation of a lone pair on an axial Z, e.g., the nitrogen lone pair on Me₂N, to an endocyclic P-O or P–N antibonding orbital or via $p\pi$ –d π bonding also could stabilize the axial P-Z bond. Because of expected steric restrictions on axial P-Z bond conformations, this sort of

⁽¹¹⁾ Haemers, M.; Ottinger, R.; Zimmermann, D.; Reisse, J. Tetra-

 ⁽¹²⁾ Kirby, A. J. The Anomeric Effect and Related Stereoelectronic
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 1983. Juaristi, E.; Gabriel, C. Tetrahedron 1992, 48, 5019-5087. For applications to six-membered rings see especially: Juaristi, E. Introduction New York, 1991, Chapters 17 and 18.

exo anomeric effect¹² is generally less important than the analogous stabilization that is available to an equatorial Z.



In three-coordinate 1,3,2-dioxaphosphorinanes, vicinal dieguatorial steric interactions are absent. Stereoelectronic effects dominate and orient nearly all substituents preferentially axial (CH₃, CH₃O, PhO, Cl, Ph).^{1,9,13} The six-membered ring is in the chair conformation like 20 (NPh = O). With more sterically demanding substituents, e.g., isopropyl and tert-butyl, 1,3-syn axial interactions increase and shift the equilibrium toward 21 (NPh = O).⁹ Substituents NMe₂ and tert-butyl strongly prefer the equatorial orientation, the latter for obvious steric reasons. The 1 kcal/mol equatorial preference of Me₂N is believed to result to a large extent from optimal stereoelectronic stabilization in the equatorial orientation that is lost in the axial position, 21.9,13 The MeNH substituent (Z) leads to an approximately equal population of 20 and 21.^{13a}

As shown above for 7–16 and by Nifant'ev et al. for 5 and 6,4ª substituents (Z) on phosphorus such as OMe, OCH-(CF₃)₂, and Ph in three-coordinate 1,3,2-oxazaphosphorinanes also strongly favor the axial orientation (20).4a,6 However, by contrast to the findings for 1,3,2-dioxaphosphorinanes containing three-coordinate phosphorus, NHMe and even NMe₂ groups are seen to be predominantly axial in the three-coordinate 1,3,2-oxazaphosphorinanes studied. One or more of the relevant steric or stereoelectronic factors mentioned above may be responsible. Firstly, a reduction in the magnitude of 1,3-syn axial repulsions in 20 can be anticipated from the X-ray structures of 11 and 13. The P-N and C-N bond lengths (Table II) are longer than those for P-O and C-O, respectively, which increases the distance between axial protons and axial NMe₂. In addition, the N-containing six-membered ring appears to be readily deformable in relief of 1,3-syn axial steric repulsions. This is seen in the bond angles P(2)N(3)C(4)(125.8°) and P(2)O(1)C(6) (123.4°) of 13, which are considerably larger than those of 11 (118.8 and 119.9°, respectively), and in the interplane angles for 11 and 13 depicted in structures 24 and 25. (Best plane through O1,



N(3), C4, C6 used). The angle P(2)N(9)C(10) is enlarged to 127.2 (2)°, perhaps also to decrease 1,3-syn axial repulsions. As a result the axial Me₂N is rotated about its P-N bond so as to move one methyl group over the ring and closer to the axial hydrogen at C4 while maintaining near planarity about the axial nitrogen (sum of angles about nitrogen, 358°), Figure 4. With an assumed van der

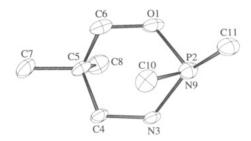


Figure 4. Truncated ORTEP plot for 13 showing conformation about the axial P-N bond.

Waals radius of 2.0 Å for the CH₃¹⁴ group and 1.20 Å for hydrogen,¹⁵ the measured C····H_C internuclear distance (3.6 Å) is considerably beyond the van der Waals sum (3.2 Å). That for C····H_A (3.1 Å) is somewhat shorter.

The second steric factor, one which is not present in 1,3,2-dioxaphosphorinanes, involves potential vicinal repulsive interactions in 20 or 21 between the substituent Z on phosphorus and the *i*-Pr (5 and 6) or Ph (7-16) substituent on ring nitrogen (N(3)). Indeed, in the corresponding 2-oxo-1,3,2-oxazaphosphorinanes, such repulsions have been shown to destabilize conformations with axial Z corresponding to 20 and cis-17a.^{2a-h} This effect appears not to be a dominant factor in the threecoordinate cases, however. As shown in other work in this laboratory, the details of which will be published separately,¹⁶ replacement of N(3)Ph in 13 or 14 by N(3)CH₃ (structure 26) shifts the equilibrium toward 21 (26b) (Table V). By contrast, the same substitution in the 2-oxo series shifts the chair-twist equilibrium for $Z = Me_2N^{17}$ in the opposite direction and has the same effect on both chairchair and chair-twist equilibria for $Z = Ph.^{2h}$ The replacement of Ph at N(3) with the much smaller hydrogen has an even greater effect in the same direction.^{2a-h} These effects have been assigned^{2a-h} to dominant vicinal axialequatorial repulsions present between axial Me₂N and Ph on phosphorus and substituents at N(3) decreasing in the order Ph > Me > H. Conversely, we interpret the greater stability of 20 over 21 in 13 and 14 to result primarily from steric repulsion accompanying the equatorial-equatorial positioning of the Me_2N and PhN(3) in 21.

We also measured the coupling constants for 4 at 300 MHz (Table III) and found them to be surprisingly close to those determined by Nifant'ev et al. from a less welldispersed 100-MHz spectrum. The percentage of the Me₂N-axial chair conformer corresponding to 20 was determined for 4 to be 77% (Table V) in the same manner as it was for 7-16 (eqs 3 and 4). Recall the result discussed earlier in which replacement of the i-Pr at N(3) or 4 with Me (26) was seen to shift the equilibrium strongly toward 21 (26b). (Indeed, about 70% of 26b becomes populated (Table V)). Thus, the N(3) substitutents, Ph and i-Pr, both of which are larger than Me, destabilize 21 and shift the equilibrium toward Me₂N-axial conformations represented by 20.

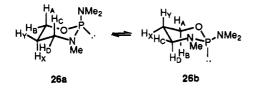
The importance of stereoelectronic effects not present in 1,3,2-dioxaphosphorinanes, a possible third factor in the enhanced preference of Me₂N and MeNH substituents on phosphorus for the axial position in 1,3,2-oxazaphos-

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phorinanes, is not so readily assessed. However, the orientation of the Me₂N in 13 (Figures 2 and 4) is such that its nitrogen lone pair can be involved in stabilizing $n-\sigma^*$ donation to the σ^* orbital of the O(1)-P bond. (The preferred orthogonality of phosphorus and nitrogen lone pairs is well known.¹⁸) This stabilization would result in a kind of exo anomeric effect.¹² In addition, the conventional endo anomeric effect may be enhanced in 1,3,2oxazaphosphorinanes if the N(3) lone pair is higher in energy than that of a lone pair on ring oxygen thus increasing overall $n-\sigma^*$ stabilization. It had been hoped that the substituents on the phenyl rings of 15 and 16 might perturb the energy of the N(3) lone pair sufficiently to alter the equilibrium of $20 \rightleftharpoons 21$. However, no perceptible change occurs. Either the perturbation is too small to be seen or $n \rightarrow \sigma^*$ interactions involving the N(3) lone pair and the axial σ_{PZ}^* orbital are not important in these molecules. The latter seems highly unlikely. Perhaps an increase in $n \rightarrow \sigma^*$ stabilization involving the equatorial phosphorus lone pair and the endocyclic C-N antibonding orbital results when an electron-withdrawing substitution is placed on the N(3)Ph. This would offset the expected increased n_N/σ_{PZ}^* stabilization when Z is axial.

Chair-Twist Equilibria. That the cis diastereomer of 17 is the more stable one is not surprising in view of the axial preference of the Me_2N in 13 and 14. Nonetheless, boat/twist forms must be very energetically available for cis-17. Thus, even more of cis-17d (Me₂N pseudoequatorial) is populated in equilibrium with cis-17a (cis-17a/ $cis-17d \simeq 60/40$) than is 21 (Me₂N equatorial) in chairchair equilibrium with 20 for 14 (20/21 = 80/20). Dreiding models show that in boat/twist form cis-17d the Me₂N and N(3)Ph are moved away from one another (P-N and N-Ph bonds less eclipsed) to relieve the equatorialequatorial repulsion that leads to the destabilization present in trans-17a (and 21). This also makes boat/twist form cis-17d of lower energy than cis-17c and explains the lack of population of the latter. It is likely the reduced equatorial-equatorial repulsion in cis-17d that makes it so readily formed from cis-17a. It is difficult to estimate quantitatively the intrinsic resistance of conformation cis-17a to conversion to twist form cis-17d. The driving force for the cis-17a \rightarrow cis-17d reorientation of the axial Me₂N cannot be estimated from the equilibrium 20 \rightleftharpoons 21 because in cis-17d there is not present the repulsive equatorialequatorial $N(3)Ph/NR_2$ interaction of 21. Nonetheless, the chair-twist free energy difference must be very small as it is in three-coordinate 1,3,2-dioxaphosphorinanes.^{9,19}

For trans-17, the destabilizing equatorial-equatorial interaction present in 21 is operative in trans-17a but relieved in formation of trans-17b and trans-17c. The

near-equal energies of trans-17b and trans-17c are reasonable in view of the pseudoaxial positions of the Me₂N in both which, therefore, are devoid of the equatorialequatorial repulsions present in twist form cis-17c and in chair conformations trans-17a and 21. As for cis-17, twist conformations must be very little higher in energy intrinsically than chair conformations. Here, the equilibria do involve, in formation of cis-17b and cis-17c, relief of the equatorial/equatorial repulsion felt in trans-17a. Since equilibrium 20 \Rightarrow 21 favors the Me₂N axial form by about 0.8-1.1 kcal/mol (13 and 14 Table V) the resistance to chair to twist interconversion to place the Me₂N pseudoaxial (trans-17b and trans-17c) must be a few tenths of a kcal greater than 0.8-1.1 kcal/mol, as trans-17a is not fully depopulated.

It also should be noted that the boat/twist conformations formed from *cis*- and *trans*-17 are not the same. Thus, *trans*-17b and -17c have the Me₂N pseudoaxial, and those formed from *cis*-17a have the Me₂N pseudoequatorial. Notably, boat/twist forms are not populated by 22,⁹ as is consistent with the propensity of its Me₂N to be equatorial.

Low free energy differences for three-coordinate 1,3,2oxazaphosphorinanes of chair, and the ease of population of boat or twist forms, are not surprising since this also is true for three- 9,19 and four-coordinate 1,3,2-dioxaphosphorinanes²⁰⁻²² and four-coordinate 1,3,2-oxazaphosphorinanes.^{2c,d,g,h} In these ring systems it is usually presumed that the relatively long endocyclic P–O and P–N bonds and accompanying flattening of the rings about phosphorus reduce not only 1,3-*syn* axial repulsions in chair forms but also cross-ring torsional repulsions in boat/ twist conformations. The very long P–N bonds of threecoordinate 1,3,2-oxazaphosphorinanes (Table II) may make boat/twist forms even more accessible.

Conclusion

The series of 1,3,2-oxazaphosphorinanes 7-11 with Z = MeO, $(CF_3)_2CHO$, or Ph at phosphorus behaves conformationally much like the well-studied 1,3,2-dioxaphosphorinanes. Each populates a single chair conformation, 20. This is most likely in response to $n \rightarrow \sigma^*$ stereoelectronic interactions related to the anomeric effect that favor axial orientation of Z to the degree that repulsive 1,3-syn axial interactions present in 20 are overcome. The ring with Z = MeNH, 12, also is essentially all in conformer 20 in solution, even though in the 1,3,2-dioxaphosphorinane

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⁽²²⁾ For rings of cyclic nucleotides and related five/six fused systems see ref 3 and: Nelson, K. A.; Bentrude, W. G. Carbohydr. Res. 1992, 234, 141-156. Sopchik, A. E.; J. Beres, J. Tomasz, Bentrude, W. G. J. Org. Chem. 1991, 56, 5911-5918. Hermans, R. J. M.; Buck, H. M. Recl. Trav. Chim. Pays-Bas 1988, 107, 82-88. Bottka, S.; Radics, L.; Tomasz, J. Nucleosides Nucleotides 1989, 8, 1217-1229.

ring system the MeNH substituent displays a nearly neutral equatorial-axial preference in solution. Moreover, the Me₂N of 13 and 14 is preferentially axial as measured by the 80-90% population of chair conformer 20 by these molecules. Thus, conformer 20 is 0.8-1.1 kcal/mol more stable than is conformer 21 in contrast to the approximate 1 kcal/mol preference of the Me₂N for the equatorial position when attached to phosphorus of the corresponding 1,3,2-dioxaphosphorinane ring. This surprising finding evidently is partially rationalized in terms of the reduction in 1,3-syn axial repulsions for axial Me_2N in the 1,3,2oxazaphosphorinane ring. Thus, in addition to the presence of relatively long endocyclic N-P bonds, evidence is seen from the X-ray structures of 11 and 13 that the ring of 13 is distorted somewhat to accommodate the axial Me_2N of 13 which is rotated about the axial P-N bond into a conformation in which it benefits from close to optimal stereoelectronic interactions. However, the previous proposal of Nifant'ev regarding axial orientation of the Me₂N of 4, which we showed above to be 75-80% in conformation 20, together with unpublished results from this laboratory concerning the primary population of 21 by 26, the analog of 14 with Me in place of Ph at N(3), requires an additional, overriding consideration. The dominant influence on the equilibrium $20 \approx 21$ for 4, 13, and 14 appears to be the strong destabilization of 21 by steric repulsions between the relatively large substituents Ph and *i*-Pr at N(3) and the equatorial Me₂N. The dominance of steric repulsions between relatively large substitutents on N(3) and equatorial Me₂N on phosphorus is in contrast to the established influence on conformations of the corresponding 2-oxo-1,3,2-oxazaphosphorinanes of repulsions between axial Me₂N or Ph on phosphorus and Me or Ph at N(3). The latter interactions were shown to destabilize the 2-oxo form of 20 in favor of the chair conformation corresponding to 21.

The essential correctness of the above ideas is further supported by their use to rationalize the conformational (chair-boat/twist) equilibria observed for cis- and trans-17. It also is evident from the studies of cis- and trans-17 that the free energy difference between chair and boat/ twist conformations of three-coordinate 1,3,2-oxazaphosphorinanes is comparatively small. These studies clearly demonstrate the variety of steric and stereoelectronic interactions that are operative in the conformational equilibria of 1,3,2-oxazaphosphorinanes. One also sees the contrasting conformational features of 1,3,2-dioxa- and 1,3,2-oxazaphosphorinanes that possess three-coordinate phosphorus, and the differences between the conformational properties of 1,3,2-oxazaphosphorinanes containing three-coordinate phosphorus and those of their 2-oxo counterparts.

Experimental Section

Materials. Commercial reagents and solvents were used as received unless otherwise noted. Diethyl ether and tetrahydrofuran (THF) were dried over sodium/benzophenone. Both were freshly distilled before use. Other solvents were OmniSolv grade from EM industries Inc. Zinc chloride was dried under vacuum with occasional heating with a heat gun for 30 min. Anilines were freshly distilled before use. 2,2-Dimethyl-3-hydroxypropionaldehyde, purchased from Aldrich Chemical Co., was purified by Kugelrohr distillation. aquisitions. ¹H NMR spectra were recorded on Varian XL-300, Unity-300, and VXR-500 spectrometers. ¹³C NMR spectra were determined at 75 MHz on Varian XL-300 and Unity-300 spectrometers. An APT (attached proton test) ¹³C NMR spectrum was obtained to aid the assignments of the resonances of 17. ¹H and ¹³C NMR chemical shifts are expressed in ppm relative to the internal tetramethylsilane (TMS). Detailed NMR parameters not given in Tables III–IV are recorded in this section. For compounds 7–17 only ¹H NMR couplings not given in the tables are recorded below.

Crystals 11 and 13, both from n-pentane, with dimensions of $0.23 \times 0.19 \times 0.15$ mm³ and $0.28 \times 0.15 \times 0.08$ mm³, respectively, were glued and sealed in capillary tubes under argon atmosphere and then mounted on the instrument. The data were collected on an Enraf-Nonius CAD4 diffractometer at -125 °C. Cell constants were obtained from 25 reflections within $15^{\circ} < 2\theta <$ 25°. The space groups were determined from subsequent leastsquares refinement. Standard reflections showed decay (4% for 11 and 12% for 13) during data collection, and an anisotropic decay correction was applied. Lorentz and polarization corrections, and an empirical absorption correction, based on a series of ψ scans, were applied to the data. The structures were solved by direct method techniques with a SDP/VAX package. All hydrogen atoms were located and refined with fixed isotropic thermal parameters. Scattering factors²³ and $\Delta f'$ and $\Delta f''$ factors²⁴ were taken from the literature. A more detailed description of the above procedures has been published.^{25,26}

Mass spectra (GC/MS and GC/HRMS) were recorded on a Finnigan MAT 95 instrument operating in the electron ionization mode (EI). Gas chromatographic analyses were performed on a Varian 3300 gas chromatograph equipped with an HP-capillary column (25 m \times 0.32 mm) and flame ionization detection. Elemental analyses were performed by Atlantic Microlab, Inc., Norcross, GA, and Galbraith laboratories, Inc., Knoxville, TN. Melting points are uncorrected.

Preparation of N-Phenyl-2-(hydroxymethyl)-2-methylpropylamine. Zinc chloride (3.65 g, 26.8 mmol) and sodium cyanoborohydride (3.43 g, 51.9 mmol) were added to 90 mL of methanol. The solution was stirred at room temperature for 1 h and then poured into a solution of 2,2-dimethyl-3-hydroxypropionaldehyde (5.47 g, 53.5 mmol) and aniline (6.86 g, 73.6 mmol) in 140 mL of methanol at room temperature which was then rapidly stirred under argon over a 36-h period. The reaction was quenched with 300 mL of 0.1 N NaOH. After most of the methanol was removed under reduced pressure, the aqueous solution was extracted with 3×150 mL of diethyl ether. The combined extracts were washed with brine and dried over $MgSO_4$. The solvent was removed under reduced pressure, and the residue was short-path distilled under vacuum to give 6.33 g of a colorless liquid (35.3 mmol, 66% yield): bp 110-114 °C (0.2 mmHg) (lit.²⁴ bp 108–110 °C (0.25 mmHg)).

Preparation of N-(m-(Trifluoromethyl)phenyl)-2-(hydroxymethyl)-2-methylpropylamine. By a procedure directly analogous to the above, reaction of zinc chloride (3.65 g, 26.8 mmol) and sodium cyanoborohydride (3.36 g, 50.9 mmol) in 90 mL of methanol and a solution of 2,2-dimethyl-3-hydroxypropionaldehyde (5.47 g, 53.5 mmol) and m-(trifluoromethyl)aniline (12.9 g, 80.3 mmol) in 140 mL of methanol at room temperature gave 7.31 g of product as a colorless liquid (29.6 mmol, 55% yield): bp 112-115 °C (0.25 mmHg) (lit.²⁷ bp 104-105 °C (0.07 mmHg)).

Preparation of 3-(Phenylamino)-1-propanol. A mixture of 3-chloro-1-propanol (17.4 g, 15.4 mL, 0.181 mol) and aniline (50 mL, 51.1 g, 0.549 mol) was heated at 95–105 °C for 5 h. The reaction mixture was taken up in 100 mL of diethyl ether and washed with 2×100 mL of 1 N NaOH. The combined aqueous

Spectral and Physical Data. ³¹P NMR spectra were taken on Varian XL-300 and Unity-300 spectrometers at 121 MHz with proton-decoupling ($\{^{1}H\}$). Positive ³¹P NMR chemical shifts are expressed in ppm downfield from external 85% H₃PO₄. Integrated ³¹P spectra (17) were obtained using a 50-s delay between

⁽²³⁾ Cromer, D. T.; Waber, J. T. International Tables for X-ray Crystallography; Kynoch Press: Birmingham, England, 1974; Vol. IX, Table 2.2B.

⁽²⁴⁾ Cromer, D. T. International Tables for X-ray Crystallography; Kynoch Press: Birmingham, England; Vol. IV, Table 2.3.1.

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⁽²⁶⁾ The authors have deposited atomic coordinates for these structures with the Cambridge Crystallographic Data Centre. The coordinates can be obtained, on request, from the Director, Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge, CB2 1EZ, UK.

layers were extracted with diethyl ether $(2 \times 100 \text{ mL})$. The combined organic layers were washed with brine and dried over MgSO₄. The solvent was removed by rotary evaporation, and the residue was short-path distilled to give 19.7 g of a thick yellow-green liquid (0.131 mol, 72% yield): bp 122-126 °C (0.3 mmHg) (lit.²⁸ bp 172-179 °C (11 mmHg)); ¹H NMR (300 MHz, CDCl₃) δ 1.87 (quintet, ³J_{HH} = 6.1 Hz, 2 H, CH₂CH₂CH₂), 3.27 (t, 2 H, CH₂NH), 3.80 (t, 2 H, CH₂OH), 6.62-6.79, 7.15-7.21 (m, 5 H, C₆H₆); ¹⁸C NMR (75 MHz, CDCl₃, {¹H}) δ 31.63 (s, 1 C, CH₂CH₂-CH₂), 41.55 (s, 1 C, CH₂NH), 6.101 (s, 1 C, CH₂OH), 112.98 (s, 2 C, o-Ph), 117.44 (s, 1 C, p-Ph), 129.08 (s, 2 C, m-Ph), 148.16 (s, 1 C, *ipso*-Ph). Anal. Calcd for C₉H₁₃NO: C, 71.49; H, 8.63; N, 9.22. Found: C, 71.23; H, 8.64; N, 9.20.

2-(Dimethylamino)-3-isopropyl-1,3,2-oxazaphosphorinane (4) was prepared by the method of Nifant'ev et al.^{4a} The requisite 2-chloro-3-isopropyl-1,3,2-oxazaphosphorinane precursor was obtained from PCl₃ and the amino alcohol, prepared by a literature procedure.²⁹

Preparation of 2-Methoxy-3-phenyl-5,5-dimethyl-1,3,2oxazaphosphorinane (7). A solution of 2-chloro-3-phenyl-5,5dimethyl-1,3,2-oxazaphosphorinane²⁴ (3.60 g, 14.8 mmol) in 25 mL of dry diethyl ether was added dropwise to a solution of methanol (0.50 g, 16.3 mmol) and triethylamine (1.70 g, 16.3 mmol) in 55 mL of dry diethyl ether at 0 °C with rapid stirring. The mixture was slowly warmed to room temperature and continuously stirred overnight. The salt was removed by Schlenk techniques, and the solvent was removed by rotary evaporation. The residue was short-path distilled to give 2.90 g of a colorless liquid, 99% pure by GC (12.0 mmol, 81% yield): bp 90-91 °C (0.025 mmHg). Further purification by molecular distillation gave analytically pure product (bp 40–41 °C (0.025 mmHg)): $^{31}\mathrm{P}$ NMR (121 MHz, C₆D₆, $\{{}^{1}H\}$) δ 125.5 (s); ${}^{1}H$ NMR (300 MHz, C_6D_6) δ 0.47, 1.09 (two s, 6 H, C(CH₃)₂), 2.66 (ddd, 1 H, H_D, ²J_{CD}) = -11.8 Hz), 3.18 (ddd, 1 H, H_B, ${}^{2}J_{AB}$ = -10.4 Hz), 3.19 (d, ${}^{3}J_{PH}$ = 12.1 Hz, 3 H, OCH₃), 3.51 (dd, 1 H, H_c, ${}^{2}J_{CD}$ = -11.8 Hz), 3.98 (dd, 1 H, H_A, ${}^{2}J_{AB} = -10.4$ Hz), 6.83–6.89, 7.13–7.15 (m, 5 H, phenyl); ${}^{13}C$ NMR (75 MHz, C₆D₆, 1 H}) δ 23.08, 24.23 (two s, 2 C, C(CH₃)₂), 31.85 (s, 1 C, C₅), 50.67 (d, 1 C, OCH₃, ${}^{2}J_{PC} = 17.7$ Hz), 54.30 (d, 1 C, C₄, ${}^{2}J_{PC} = 2.1$ Hz), 68.35 (d, 1 C, C₆, ${}^{2}J_{PC} = 2.1$ Hz), 20.8 (d, 1 C, C P, B 2.8 Hz), 120.69 (d, 2 C, o-Ph, ${}^{3}J_{PC} = 13.1$ Hz), 122.08 (d, 1 C, p-Ph, ${}_{\rm SJ_{PC}}$ = 2.1 Hz), 129.38 (s, 2 C, m-Ph), 148.34 (d, 1 C, ipso-Ph, $_{2J_{PC}}$ = 17.9 Hz). Anal. Calcd for C₁₂H₁₈NO₂P: C, 60.24; H, 7.58; N, 5.85. Found: C, 60.18; H, 7.61; N, 5.79.

Preparation of 2-(1,1,1,3,3,3-Hexafluoroisopropoxy)-3phenyl-5,5-dimethyl-1,3,2-oxazaphosphorinane (8). By a procedure directly analogous to that for the preparation of 7, the reaction of 1,1,1,3,3,3-hexafluoro-2-propanol (2.50 g, 14.4 mmol) and triethylamine (1.50 g, 14.4 mmol) in 50 mL of dry diethyl ether and 2-chloro-3-phenyl-5,5-dimethyl-1,3,2-oxazaphosphorinane²⁴ (3.50 g, 14.4 mmol) in 20 mL of dry diethyl ether gave 4.50 g of a colorless liquid (12.0 mmol, 83% yield): bp 82-83 °C (0.025 mmHg); ³¹P NMR (121 MHz, C₆D₆, {¹H}) δ 131.1 (septet, ⁴J_{PF} = 6.2 Hz); ¹H NMR (300 MHz, C₆D₆) δ 0.34, 0.97 (two s, 6 H, $C(CH_3)_2$, 2.48 (ddd, 1 H, H_D, ${}^2J_{CD} = -11.9$ Hz), 3.17 (ddd, 1 H, H_B , ${}^2J_{AB} = -10.6 Hz$), 3.41 (dd, 1 H, H_C, ${}^2J_{CD} = -11.9 Hz$), 3.97 $(dd, 1 H, H_A, {}^2J_{AB} = -10.6 Hz), 4.04 (d of septet, 1 H, OCH(CF_3)),$ ${}^{3}J_{\rm PH} = 13.6$ Hz, ${}^{3}J_{\rm FH} = 6.0$ Hz), 6.87-6.93, 7.00-7.04, 7.08-7.14 (m, 5 H, phenyl); ¹³C NMR (75 MHz, C₆D₆, {¹H}) δ 22.67, 23.76 $(two s, 2 C, C(CH_3)_2), 31.36 (d, 1 C, C_5, {}^{3}J_{PC5} = 1.0 Hz), 55.28 (d, 1 C, C_5, {}^{3}J_{PC5} = 1.0$ 1 C, C₄, ${}^{2}J_{PC4} = 2.7$ Hz), 69.68 (d, 1 C, C₆, ${}^{2}J_{PC6} = 3.6$ Hz), 71.01 (d of septet, 1 C, OCH(CF₃)₂, ${}^{2}J_{PC} = 21.0$ Hz, ${}^{1}J_{FC} = 33.4$ Hz), 122.29 (d, 2 C, o-Ph, ${}^{3}J_{PC}$ = 11.4 Hz), 123.87 (d, 1 C, p-Ph, ${}^{5}J_{PC}$ = 2.5 Hz), 129.49 (s, 2 C, m-Ph), 146.37 (d, 1 C, *ipso*-Ph, ${}^{2}J_{PC}$ = 18.4 Hz). The CF_3 carbon signals were too weak to be observed. Anal. Calcd for C14H16F6NO2P: C, 44.81; H, 4.30; N, 3.73. Found: C, 44.87; H, 4.31; N, 3.67.

Preparation of 2-Methoxy-3-phenyl-1,3,2-oxazaphosphorinane (9). By a procedure directly analogous to that for the preparation of 7, the reaction of methanol (0.71 g, 0.90 mL, 22.0 mmol) and triethylamine (2.22 g, 22.0 mmol) in 50 mL of dry diethyl ether and 2-chloro-3-phenyl-1,3,2-oxazaphosphorinane²⁴ (4.31 g, 20.0 mmol) in 150 mL of dry diethyl ether gave 3.55 g of a colorless liquid (16.8 mmol, 84% yield): bp 90 °C (0.025 mmHg); ³¹P NMR (121 MHz, C₆D₆, {¹H}) δ 133.7 (s); ¹H NMR (300 MHz, C₆D₆) δ 1.15 (dddd, 1 H, H_Y, ²J_{XY} = -13.7 Hz), 1.90 (dddd, 1 H, H_X, ²J_{XY} = -13.7 Hz), 2.88 (ddddd, 1 H, H_D, ²J_{CD} = -12.0 Hz), 3.19 (d, 3 H, OCH₃, ³J_{PH} = 12.2 Hz), 3.50 (dddd, 1 H, H_B, ²J_{AB} = -10.7 Hz), 3.57 (dddd, 1 H, H_C, ²J_{CD} = -12.0 Hz), 4.07 (dddd, 1 H, H_A, ²J_{AB} = -10.7 Hz), 6.83–6.90, 7.10–7.17 (m, 5 H, phenyl); ¹³C NMR (75 MHz, C₆D₆ {¹H}) δ 27.19 (d, 1 C, C₅, ³J_{PC5} = 1.7 Hz), 42.60 (d, 1 C, C₄, ²J_{PC4} = 2.0 Hz), 50.68 (d, ²J_{PC} = 17.9 Hz, 1 C, OCH₃), 59.06 (d, 1 C, C₆, ²J_{PC6} = 2.7 Hz), 119.98 (d, 2 C, o-Ph, ³J_{PC} = 13.3 Hz), 121.85 (d, 1 C, p-Ph, ³J_{PC} = 15.9 Hz), 129.33 (s, 2 C, m-Ph), 148.06 (d, 1 C, *ipso*-Ph, ²J_{PC} = 17.9 Hz). Anal. Calcd for C₁₀H₁₄NO₂P: C, 56.87; H, 6.68; N, 6.63. Found: C, 56.81; H, 6.69; N, 6.58.

Preparation of 2-(1,1,1,3,3,3-Hexafluoroisopropoxy)-3phenyl-1,3,2-oxazaphosphorinane (10). By a procedure analogous to that for the preparation of 7, the reaction of 2-chloro-3-phenyl-1,3,2-oxazaphosphorinane²⁴ (6.73 g, 31.2 mmol) in 150 mL of diethyl ether with triethylamine (3.47 g, 4.80 mL, 34.3 mmol) and hexafluoro-2-propanol (5.83 g, 3.7 mL, 34.3 mmol) in $50~\mathrm{mL}$ of diethyl ether gave 9.91 g of product (28.5 mmol, 91 \% yield): bp 97-98 °C (0.25 mmHg); ³¹P NMR (121 MHz, C₆D₆, ${}^{1}H_{}$ δ 138.7 (septet, ${}^{4}J_{PF} = 6.2$ Hz); ${}^{1}H$ NMR (300 MHz, C₆D₆) δ 1.08 (ddddd, 1 H, H_y, ²J_{xy} = -14.0 Hz), 1.68 (ddddd, 1 H, H_x, ${}^{2}J_{XY} = -14.0$ Hz), 2.69 (ddddd, 1 H, H_D, ${}^{2}J_{CD} = -11.7$ Hz), 3.38 $(dddd, 1 H, H_C, {}^{2}J_{CD} = -11.7 H_Z), 3.45 (ddddd, 1 H, H_B, {}^{2}J_{AB} =$ -11.0 Hz), 4.00 (dddd, 1 H, H_A, ${}^{2}J_{AB} = -11.0$ Hz), 4.05 (d of septet, 1 H, OCH(CF₃)₂, ${}^{3}J_{PH} = 7.7$ Hz, ${}^{3}J_{FH} = 6.0$ Hz), 6.85–6.91, 6.77– 7.01, 6.97-7.01, 7.08-7.14 (m, 5 H, phenyl); ¹³C NMR (75 MHz, C_6D_6 {¹H}) δ 26.36 (d, 1 C, C_5 , $^{3}J_{PC5} = 2.1$ Hz), 43.44 (d, 1 C, C_4 , $^{2}J_{PC4} = 2.8$ Hz), 60.56 (d, 1 C, C_6 , $^{2}J_{PC6} = 3.6$ Hz), 70.01 (d of septet, 1 C, OCH(CF₃)₂, $^{2}J_{PC} = 21.1$ Hz, $^{2}J_{PC} = 33.2$ Hz), 121.30 (d, 2 C, o-Ph, ${}^{3}J_{PC} = 12.0$ Hz), 122.08 (q, 2 C, CH(CF₃)₂, ${}^{1}J_{FC} =$ 284.2 Hz), 123.54 (d, 1 C, p-Ph, ${}^{5}J_{PC} = 2.6$ Hz), 129.43 (s, 2 C, m-Ph), 146.18 (d, 1 C, *ipso*-Ph, ${}^{2}J_{PC} = 18.2$ Hz). Anal. Calcd for C12H12F6NO2P: C, 41.51; H, 3.48; P, 8.92; N, 4.03. Found: C, 41.48; H, 3.49; P, 8.49; N, 4.04.

Preparation of 2,3-Diphenyl-5,5-dimethyl-1,3,2-oxazaphosphorinane (11). A solution of N-phenyl-2-(hydroxymethyl)-2-methylpropylamine (2.79 g, 15.6 mmol) and bis(diethylamino)phenylphosphine³⁰ (3.93 g, 15.6 mmol) in refluxing acetonitrile (250 mL) over a period of 9 days gave 3.00 g of product as an oil (10.5 mmol, 67% yield) which became solid at -20 °C and was recrystallized from *n*-pentane to give white crystals: mp 56.5-57.0 °C; ³¹P NMR (121 MHz, C₆D₆, {¹H }) δ 111.9 (s); ¹H NMR (300 MHz, C₆D₆) δ 0.17, 1.08 (two s, 6 H, C(CH₃)₂), 3.06 (ddd, 1 H, H_D, ${}^{2}J_{CD} = -13.6$ Hz), 3.19 (d, 1 H, H_C, ${}^{2}J_{CD} = -13.6$ Hz), 3.28 (ddd, 1 H, H_B, ${}^{2}J_{AB} = -10.7$ Hz), 3.61 (dd, 1 H, H_A, ${}^{2}J_{AB}$ = -10.7 Hz), 6.84-6.89, 7.05-7.10, 7.13-7.22, 7.46-7.51 (m, 10 H, two phenyl); ¹³C NMR (75 MHz, CDCl₃, {¹H}) δ 23.13, 23.69 (two s, 2 C, C($(CH_3)_2$), 33.30 (d, 1 C, C₅, ${}^{3}J_{PC5} = 1.0$ Hz), 50.03 (d, 1 C, C₄, ${}^{2}J_{PC4} = 3.1$ Hz), 72.23 (d, 1 C, C₆, ${}^{2}J_{PC6} = 5.4$ Hz), 119.26 (d, 1 C, C₆, {}^{2}J_{PC6} = 5.4 Hz), 119.26 (d, 1 C, C₆, {}^{2}J_{PC6} = 5.4 Hz), 119.26 (d, 1 C, C₆, {}^{2}J_{PC6} = 5.4 Hz), 119.26 (d, 1 C, C_{6} = 5.4 Hz), 119.26 (d, 1 C, C_{6} = 5.4 Hz), 2 C, o-PhN, ${}^{3}J_{PC} = 15.8$ Hz), 120.98 (d, 1 C, p-PhN, ${}^{5}J_{PC} = 1.7$ Hz), 129.47 (s, 2 C, m-PhN), 150.92 (d, 1 C, ipso-PhN, ²J_{PC} = 23.8 Hz), 128.83 (d, 1 C, p-PhP, ${}^{4}J_{PC} = 1.7$ Hz), 129.21 (d, 2 C, o-PhP, ${}^{2}J_{PC} = 2.0$ Hz), 130.23 (d, 2 C, m-PhP, ${}^{3}J_{PC} = 15.6$ Hz), 140.74 (d, 1 C, ipso-PhP, ${}^{1}J_{PC} = 30.7$ Hz). Anal. Calcd for C₁₇H₂₀NOP: C, 71.56; H, 7.07. Found: C, 71.64; H, 7.12.

Preparation of 2-(Methylamino)-3-phenyl-5,5-dimethyl-1,3,2-oxazaphosphorinane (12). To a rapidly stirred solution of triethylamine (2.51 g, 24.8 mmol) in 150 mL of dry diethyl ether cooled with an ice/salt bath at -15 to -12 °C was added dropwise a solution of 2-chloro-3-phenyl-5,5-dimethyl-1,3,2oxazaphosphorinane^{2g} (6.05 g, 24.8 mmol) in 50 mL of dry diethyl ether. Methylamine gas was simultaneously bubbled through the stirred solution. The admission of methylamine gas was continued for another 5 min after the addition was complete. The resulting mixture was slowly warmed to room temperature and continuously stirred overnight. The salt was removed by Schlenk techniques, and the solvent was removed by rotary evaporation to give 4.50 g of product as an oil, >98% pure by GC (18.5 mmol, 75% yield). An attempt to further purify a sample

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by vacuum distillation failed (oil bath 180 °C (0.025 mmHg); sample was decomposed): GC/HRMS calcd for $C_{12}H_{19}N_2OP$ (m/ z) 238.1235, found 238.1240; ³¹P NMR (121 MHz, C₆D₆, {¹H}) δ 108.6 (s); ¹H NMR (300 MHz, C₆D₆) δ 0.53, 1.07 (two s, 6 H, C(CH₃)₂), 2.11 (1 H, NHCH₃, broad s), 2.33 (dd, 3 H, NHCH₃, ³J_{PH} = 12.1 Hz, ³J_{HH} = 5.9 Hz), 2.80 (ddd, 1 H, H_D, ²J_{CD} = -12.3 Hz), 2.98 (dd, 1 H, Hc, ²J_{CD} = -12.3 Hz), 3.16 (ddd, 1 H, H_B, ²J_{AB} = -10.7 Hz), 3.64 (dd, 1 H, H_A, ²J_{AB} = -10.7 Hz), 6.80-6.86, 7.14-7.16 (m, 5 H, phenyl); ¹³C NMR (75 MHz, C₆D₆, {¹H}) δ 23.12, 24.20 (two s, 2 C, C(CH₃)₂), 28.54 (d, 1 C, NHCH₃, ²J_{PC} = 24.7 Hz), 31.96 (s, 1 C, C₆), 54.65 (d, 1 C, C₄, ²J_{PC4} = 2.9 Hz), 67.57 (d, 1 C, C₆, ²J_{PC6} = 3.4 Hz), 119.50 (d, 2 C, o-Ph, ³J_{PC} = 14.4 Hz), 120.83 (d, 1 C, p-Ph, ⁵J_{PC} = 2.3 Hz), 129.25 (s, 2 C, m-Ph), 149.43 (d, 1 C, *ipso*-Ph, ²J_{PC} = 16.9 Hz).

Preparation of 2-(Dimethylamino)-3-phenyl-5,5-dimethyl-1,3,2-oxazaphosphorinane (13). A solution of N-phenyl-?-(hydroxymethyl)-2-methylpropylamine (0.99 g, 5.5 mmol) and hexamethylphosphorous triamide (HMPT) (0.90 g, 1.0 mL, 5.5 mmol) in 20 mL of dry acetonitrile was refluxed under an argon atmosphere over a period of 34 h. GC analysis showed the reaction to be complete. The solvent was removed under reduced pressure, and the residue was short-path distilled to give 0.58 g of product (2.3 mmol, 42% yield): bp 97-98 °C (0.05 mmHg). No attempt was made to optimize the yield. The product became solid at -20 °C and was recrystallized from *n*-pentane to give white crystals: mp 46-47 °C; ³¹P NMR (121 MHz, C₆D₆, {¹H}) δ 120.7 (s); ¹H NMR (300 MHz, C_6D_6) δ 0.61, 0.98 (two s, 6 H, $C(CH_3)_2$), 2.36 (d, 6 H, N(CH₃)₂, ${}^{3}J_{PH} = 8.6$ Hz), 2.92 (ddd, 1 H, H_D, ${}^{2}J_{CD}$ = -12.3 Hz), 3.29 (dd, 1 H, H_C, ${}^{2}J_{CD}$ = -12.3 Hz), 3.25 (ddd, 1 H, H_B, ${}^{2}J_{AB}$ = -10.6 Hz), 3.80 (dd, 1 H, H_A, ${}^{2}J_{AB}$ = -10.6 Hz), 6.82-6.88 (m, 5 H, phenyl); ¹³C NMR (75 MHz, CDCl₃, {¹H}) δ 23.68, 24.48 (two s, 2 C, C(CH₃)₂), 37.28 (d, 2 C, N(CH₃)₂, ${}^{2}J_{PC} = 17.9$ Hz), 32.41 (d, 1 C, C₅, ${}^{3}J_{PC} = 1.0$ Hz), 56.13 (d, 1 C, C₄, ${}^{2}J_{PC} = 0.8$ Hz), 70.41 (d, 1 C, C₆, ${}^{2}J_{PC} = 2.4$ Hz), 119.30 (d, 2 C, o-Ph, ${}^{3}J_{PC} = 14.1$ Hz), 120.84 (d, 1 C, p-Ph, ${}^{5}J_{PC} = 1.9$ Hz), 129.24 (s, 2 C, m-Ph), 149.66 (d, 1 C, *ipso*-Ph, ${}^{2}J_{PC} = 17.4$ Hz). Anal. Calcolater the term of for C₁₃H₂₁N₂OP: C, 61.89; H, 8.39; P, 12.28. Found: C, 61.77; H, 8.43; P, 12.35.

Preparation of 2-(Dimethylamino)-3-phenyl-1,3,2-oxazaphosphorinane (14). By a procedure directly analogous to that for the preparation of 13, the reaction of 3-(phenylamino)-1propanol (9.58 g, 63.4 mmol) and HMPT (10.3 g, 11.5 mL, 63.4 mmol) in ethyl acetate (80 mL) and toluene (80 mL) over a period of 23 h gave 9.39 g of a colorless liquid (41.9 mmol, 66% yield): bp 115-122 °C (0.1 mmHg); ³¹P NMR (121 MHz, C₆D₆, {¹H}) δ 126.9 (s); ¹H NMR (300 MHz, C₆D₆) δ 1.24 (ddddd, 1 H, H_Y, ²J_{XY} = -13.4 Hz), 1.69 (ddddd, 1 H, H_X, ${}^{2}J_{XY}$ = -13.4 Hz) 2.40 (d, 6 H, N(CH₃)₂, ${}^{3}J_{PH} = 8.6$ Hz), 3.12 (ddddd, 1 H, H_D, ${}^{2}J_{CD} = -12.4$ Hz), 3.39 (dddd, 1 H, H_C, ${}^{2}J_{CD} = -12.4$ Hz), 3.56 (ddddd, 1 H, H_B, ${}^{2}J_{AB} = -10.8$ Hz), 3.87 (dddd, 1 H, H_A, ${}^{2}J_{AB} = -10.8$ Hz), 6.83-6.89, 7.09-7.21 (m, 5 H, phenyl); ¹³C NMR (75 MHz, C₆D₆ {¹H}) δ 27.10 (d, 1 C, C₅, ${}^{3}J_{PC5} = 2.3$ Hz), 37.08 (d, 2 C, N(CH₃)₂, ${}^{2}J_{PC} = 18.1$ Hz), 43.97 (d, 1 C, C₄, ${}^{2}J_{PC4} = 1.1$ Hz), 60.38 (d, 1 C, C₆, ${}^{2}J_{PC4} = 2.3$ Hz), 118.61 (d, 2 C, o-Ph, ${}^{3}J_{PC} = 13.5$ Hz), 120.65 (d, 1.2), 120.61 (d, 2.2), 0.2) (d, 1.2), 0.2) 1 C, p-Ph, ${}^{5}J_{PC}$ = 1.9 Hz), 129.20 (s, 2 C, m-Ph), 149.32 (d, 1 C *ipso*-Ph, ${}^{2}J_{PC} = 17.2$ Hz). Anal. Calcd for C₁₁H₁₇N₂OP: C, 58.92; H, 7.64; N, 12.49. Found: C, 59.01; H, 7.68; N, 12.42.

Preparation of 2-(Dimethylamino)-3-(*m*-(trifluoromethyl)phenyl)-5,5-dimethyl-1,3,2-oxazaphosphorinane (15). A procedure analogous to that for the preparation of 13 was used for the reaction of *N*-(*m*-(trifluoromethyl)phenyl)-2-(hydroxymethyl)-2-methylpropylamine (9.15 g, 37.0 mmol) and HMPT (6.03 g, 6.7 mL, 37.0 mmol) in 60 mL of toluene and 60 mL of ethyl acetate for 2 days to give 8.24 g of a colorless liquid (26.9 mmol, 73% yield): bp 109-114 °C (0.05 mmHg); ³¹P NMR (121 MHz, C₆D₆, ¹H}) δ 120.8 (s); ¹H NMR (300 MHz, C₆D₆) δ 0.57, 0.86 (two s, 6 H, C(CH₃)₂, 2.30 (d, 6 H, N(CH₃)₂, ³J_{PH} = 8.6 Hz), 2.82 (ddd, 1 H, H_D, ²J_{CD} = -12.2 Hz), 3.12 (dd, 1 H, H_C, ²J_{CD} = -12.2 Hz), 3.22 (ddd, 1 H, H_B, ²J_{AB} = -10.5 Hz), 3.65 (dd, 1 H, H_A, ²J_{AB} = -10.5 Hz), 6.95 (t, 1 H, *m*-Ph, ³J_{HH} = 7.8 Hz), 7.04 (d, 1 H, *p*-Ph, ³J_{HH} = 7.8 Hz), 7.19 (dq, 1 H, *p*-Ph, ³J_{HH} = 7.8 MIZ, 7.74 (d, 1 C, C₆, ³J_{PC} = 1.1 Hz), 37.10 (d, 2 C, N(CH₃)₂, ³J_{PC} = 18.1 Hz), 56.14 (s, 1 C, C₄), 70.94 (d, 1 C, C₆, ²J_{PC} = 1.9 Hz), 114.6 (dq, 1 C, o-Ph, ²J_{PC} = 13.4 Hz, ³J_{FC} = 4.0 Hz), 116.6 (dq, 1 C, p-Ph, ⁵J_{PC}

= 1.9 Hz, ${}^{3}J_{FC}$ = 7.7 Hz), 121.4 (dq, 1 C, o-Ph, ${}^{3}J_{PC}$ = 15.3 Hz, ${}^{5}J_{FC}$ = 1.2 Hz), 124.2 (q, 1 C, CF₃, ${}^{1}J_{FC}$ = 272.4 Hz), 129.3 (s, 1 C, m-Ph), 131.1 (q, 1 C, m-Ph, ${}^{2}J_{FC}$ = 32.0 Hz), 149.5 (d, 1 C, *ipso*-Ph, ${}^{2}J_{PC}$ = 16.7 Hz). Anal. Calcd for C₁₄H₂₀F₃N₂OP: C, 52.50; H, 6.29; N, 8.75. Found: C, 52.49; H, 6.28; N, 8.67.

Preparation of 2-(Dimethylamino)-3-(p-N,N-dimethylanilino)-5,5-dimethyl-1,3,2-oxazaphosphorinane (16). A solution of N-(p-(N,N-dimethylamino)phenyl)-2-(hydroxymethyl)-2-methylpropylamine^{2j} (1.94 g, 8.72 mmol) and HMPT (1.42 g, 8.72 mmol) in 50 mL of acetonitrile was refluxed overnight. The solvent was removed under reduced pressure, and the residue was Kugelrohr distilled to give 1.20 g of a white solid (4.1 mmol, 48% yield): air bath temperature 125-130 °C (0.025 mmHg); mp 54-55 °C; ³¹P NMR (121 MHz, C₆D₆, {¹H}) δ 124.3 (s); ¹H NMR (300 MHz, C₆D₆) δ 0.73, 1.03 (two s, 6 H, C(CH₃)₂), 2.48 (d, 6 H, PN(CH₃)₂, ${}^{3}J_{PH} = 8.5$ Hz), 2.55 (s, 6 H, C₆H₄N(CH₃)₂), 2.98 (ddd, 1 H, H_D, ${}^{2}J_{CD} = -12.1$ Hz), 3.36 (ddd, 1 H, H_B, ${}^{2}J_{AB}$ = -10.5 Hz), 3.38 (dd, 1 H, H_c, ${}^{2}J_{CD}$ = -12.1 Hz), 3.85 (dd, 1 H, H_{A} , ${}^{2}J_{AB} = -10.5 Hz$), 6.65, 7.18 (m, 4 H, C₆H₄); ¹³C NMR (75 MHz, C₆D₆, {¹H}) δ 23.85, 24.52 (two s, 2 C, C(CH₃)₂), 32.58 (d, 1 C, C₅, ${}^{3}J_{PC} = 1.4$ Hz), 37.35 (d, 2 C, PN(CH₃)₂, ${}^{2}J_{PC} = 18.0$ Hz), 41.22 (s, 2 C, C₆H₄N(CH₃)₂), 58.33 (s, 1 C, C₄), 70.76 (d, 1 C, C₆, ${}^{2}J_{PC} = 1.5 \text{ Hz}$, 114.46 (s, 2 C, m-C₆H₄N(CH₃)₂), 122.60 (d, 2 C, $o-C_6H_4N(CH_3)_2$, ${}^3J_{PC} = 10.5$ Hz), 140.31 (d, 1 C, *ipso-C_6H_4N-* $(CH_3)_2$, ${}^2J_{PC} = 17.3 Hz$), 146.77 (d, 1 C, $p-C_6H_4N(CH_3)_2$, ${}^5J_{PC} =$ 1.7 Hz). The analytical sample was prepared by molecular distillation and transferred in a glove bag under a nitrogen atmosphere to an ampoule which was flame sealed. Anal. Calcd for $C_{15}H_{26}N_3OP$: C, 60.99; H, 8.87, N, 14.23. Found: C, 60.74; H, 8.81; N, 14.08

Preparation of 2-(Dimethylamino)-3-phenyl-5-tert-butyl-1,3,2-oxazaphosphorinane (17). By a procedure directly analogous to that for the preparation of 12, the reaction of 2-chloro-3-phenyl-5-tert-butyl-1,3,2-oxazaphosphorinane^{2c} (7.83 g, 28.9 mmol) and triethylamine (2.93 g, 28.9 mmol) and dimethylamine in 300 mL of dry diethyl ether for 4 h was run. The salt was removed by Schlenk techniques, and the solvent was removed in vacuo at 0 °C to give 7.46 g of a white solid product containing cis/trans diastereomers in an 18/82 (cis/trans) ratio (⁸¹P NMR) (26.6 mmol, 92% yield). After distillation the cis/trans ratio changed to 64/36. This ratio slowly changed to a stable value of 80/20 (cis/trans) in C₆D₆ at room temperature: ³¹P NMR (121) MHz, C₆D₆, $\{^{1}H\}$) δ cis-diastereomer 124.2 (s), trans-diastereomer 127.0 (s); ¹H NMR (300 MHz, C_6D_6) cis-diastereomer δ 0.690 (s, 9 H, C(CH₃)₃), 1.72 (dddd, 1 H, H_x), 2.48 (d, ${}^{3}J_{PH} = 8.8$ Hz, 6 H, $N(CH_3)_2$, 3.44 (dddd, 1 H, H_D, ${}^2J_{CD} = -12.7$ Hz), 3.52 (ddd, 1 H, H_{C} , ${}^{2}J_{CD} = -12.7 Hz$, 3.77 (dddd, 1 H, H_{B} , ${}^{2}J_{AB} = -11.2 Hz$), 3.90 (ddd, 1 H, H_A, ${}^{2}J_{AB} = -11.2$ Hz), 6.85–6.90, 7.09–7.12, 7.18–7.24 (m, 5 H, phenyl); trans-diastereomer δ 0.687 (s, 9 H, C(CH₈)₃), 1.93 (dddd, 1 H, H_X), 2.57 (d, ${}^{3}J_{PH} = 8.3$ Hz, 6 H, N(CH₃)₂), 3.26 $(ddd, 1 H, H_C, {}^2J_{CD} = -11.9 Hz), 3.50 (dddd, 1 H, H_D, {}^2J_{CD} = -11.9$ Hz), 3.73 (ddd, 1 H, H_A, ${}^{2}J_{AB} = -11.3$ Hz), 3.96 (dddd, 1 H, H_B, ${}^{2}J_{AB} = -11.3$ Hz), 6.85–6.90, 7.09–7.12, 7.18–7.24 (m, 5 H, phenyl); ¹³C NMR (75 MHz, C₆D₆, {¹H}) cis-diastereomer δ 27.17 (s, 3 C, $C(CH_3)_3$, 31.73 (s, 1 C, $C(CH_3)_3$), 36.92 (d, 2 C, $N(CH_3)_2$, ${}^2J_{PC} =$ 18.4 Hz), 45.77 (d, 1 C, C₅, ${}^{3}J_{PC5} = 1.4$ Hz), 44.87 (d, 1 C, C₄, ${}^{2}J_{PC4}$ = 2.4 Hz), 61.83 (s, 1 C, C₆), 118.22 (d, 2 C, o-Ph, ${}^{3}J_{PC}$ = 14.0 Hz), 120.43 (d, 1 C, p-Ph, ${}^{5}J_{PC} = 1.7$ Hz), 129.33 (s, 2 C, m-Ph), 149.28 (d, 1 C, *ipso*-Ph, ${}^{2}J_{PC}$ = 18.1 Hz); trans-diastereomer δ 27.29 (s, 3 C, C(CH₃)₃), 32.00 (s, 1 C, C(CH₃)₃), 36.63 (d, 2 C, N(CH₃)₂, ${}^{2}J_{PC} = 19.2 \text{ Hz}$), 46.84 (d, 1 C, C₅, ${}^{3}J_{PC5} = 3.8 \text{ Hz}$), 46.60 (d, 1 C, C₄, ${}^{2}J_{PC4} = 3.6 \text{ Hz}$), 62.88 (s, 1 C, C₆), 119.16 (d, 2 C, o-Ph, ${}^{3}J_{PC}$ = 10.1 Hz), 120.79 (d, 1 C, p-Ph, ${}^{5}J_{PC}$ = 1.4 Hz), 129.27 (s, 2 C, m-Ph), 149.25 (d, 1 C, ipso-Ph, ²J_{PC} = 14.8 Hz). Anal. Calcd for $C_{15}H_{25}N_2OP$: C, 64.26; H, 8.99; N, 9.99. Found: C, 64.10; H, 8.97; N, 9.90.

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